PI3K δ and PI3K γ as Targets for Autoimmune and Inflammatory Diseases

Timothy D. Cushing,^{*,†} Daniela P. Metz,[‡] Douglas A. Whittington,[§] and Lawrence R. McGee[†]

[†]Therapeutic Discovery, Amgen Inc., 1120 Veterans Boulevard, South San Francisco, California 94080, United States [‡]Inflammation Research, Amgen Inc., One Amgen Center Drive, Thousand Oaks, California 91320, United States [§]Molecular Structure and Characterization, Amgen Inc., 360 Binney Street, Cambridge, Massachusetts 02142, United States

■ INTRODUCTION

Autoimmune and inflammatory diseases such as rheumatoid arthritis (RA), systemic lupus erythematosus (SLE), and asthma are chronic and often progressive diseases associated with a dysregulated or an overactive immune system, respectively. The causes and the drivers of these diseases remain ill-defined. They are characterized by complex cellular interactions between multiple inflammatory cells of the innate and adaptive immune system. The approval of rituximab (Rituxan) for RA and belimumab (Benlysta) for SLE indicates an important role for B cells in the pathology of immunemediated diseases. However, the heterogeneity and complexity of the disease etiology of these conditions make the search for new good cellular targets challenging, as it is unclear who in the cellular infiltrate is a primary player of the pathology versus an "innocent" bystander. Therefore, targeting signaling molecules that are required for the activation of multiple immune cells may be the more likely route to success in combating these chronic, immune cell-mediated diseases. This review will highlight two such signaling molecules, the related lipid kinases, class I PI3K (phosphoinositide 3-kinase) δ and γ , both of which are primarily expressed in leukocytes and seem to play important roles in their function and activation.

This review will compare and contrast these two kinases as targets for autoimmune and inflammatory diseases and summarize efforts toward the development of small molecule inhibitors of PI3K δ and PI3K γ .¹⁻³

Phosphoinositides (PIs) act as second messengers in a wide variety of cellular roles including signal transduction, control of membrane trafficking and transport, cytoskeleton organization, cell survival and death, among many other functions. The target of PI3K kinase activity is the phosphoinositides within cell membranes. Phosphoinositides are incorporated within the lipid bilayer of the membrane via two fatty acids that are attached to the cytosolic inositol ring via a glycerol phosphate linker. Cytosolic PI3K phosphorylates the inositol ring at the 3position, converting PIP2 to PIP3⁴ (Figure 1). It is through the phosphorylation of phosphoinositides that class I PI3Ks control growth, survival, and proliferation of the cells. PIP3 serves as the membrane anchoring site for multiple protein kinases⁵ that contain pleckstrin homology (PH) domains, such as the serine/ threonine kinases AKT (or PKB), PDK1, and the tyrosine kinases of the Tec family, BTK, ETK, and ITK.⁶ Membrane bound AKT activates further downstream proteins such as mTOR, FOXO3a, and others. Calcium mobilization and gene transcription are dependent on the activity of Tec family members and other PIP3 binding effectors control cytoskeleton organization, membrane trafficking, and signaling efficiency. The phosphatase PTEN returns PIP3 to its precursor PIP2, serving as a regulator of PI3K activity.

PI3Ks are divided into three classes (I, II, III) based on functional and sequence homology. Although there is increasing interest in the class II and class III PI3Ks and their products, the majority of pharmaceutical research and the focus of this review are on the class I PI3Ks. Class I PI3Ks are further divided based on signaling pathways and regulatory proteins into class IA and class IB. The class IA PI3Ks comprise three closely related kinases, PI3K α , PI3K β , and PI3K δ , which exist as heterodimers composed of a catalytic subunit (p110 α , p110 β , or p110 δ) and a regulatory subunit. These respond to signaling generally through receptor tyrosine kinases (RTKs). The class IB PI3Ky signals through G-protein-coupled receptors (GPCRs) and is composed of a p110y catalytic domain that can associate with regulatory subunits distinct from the class IA isoforms. Expression of the PI3K α and PI3K β isoforms is ubiquitous, while the expression pattern of PI3K δ and PI3Ky seems more restricted, with both isoforms found primarily in leukocytes. The relatively restricted expression pattern of PI3K δ and PI3K γ , in addition to data accumulated from studies in mice, where PI3K δ and/or PI3K γ was either genetically or pharmacologically inactivated, suggests that these two isoforms play a major role in the adaptive and innate immune systems, respectively.

SMALL MOLECULE TOOLS FOR ELUCIDATING THE ROLE OF VARIOUS PI3K ISOFORMS

Two broad spectrum oral inhibitors, 1 (wortmannin)¹⁷ and 2 (LY294002)¹⁸ (Figure 2, Table 2), have been used to study the role of PI3K in various functions and disease states. With the identification of PI3K isoforms and the development of specific activity assays, inhibitors with varying selectivity profiles for one isoform over another have been discovered. Select compounds that have served as tool molecules to elucidate the role of PI3K isoforms are listed in Figure 3 and Table 2 along with their relative potencies against the various isoforms. Inhibitors with 60- to 1000-fold selectivity for PI3K δ over the other three class I isoforms are 3 (IC87114)¹⁹ and 4 (PIK-39).¹⁹ Compounds have also been discovered that are selective for PI3K γ such as 5 (AS-252424)²⁰ and 6 (AS-605240).²¹ In addition, profiles that can be characterized as dual activity are observed for compounds such as 7 (TG100-115).²² A challenge of drug

ACS Publications © 2012 American Chemical Society

Received:
 June 15, 2012

 Published:
 August 27, 2012

Perspective

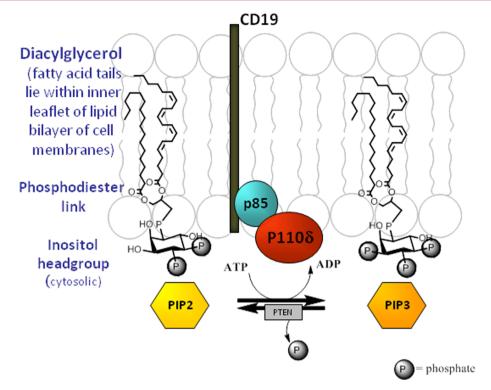
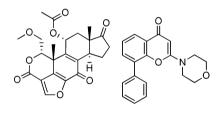


Figure 1. The function of PI3K (p85 regulatory, p110 catalytic domain) is to phosphorylate PIP2, converting it to the phosphoinositide PIP3. The phosphatase PTEN reverses the process. There are four types of class I PI3Ks: class IA, PI3K α , - β , - δ ; class 1B, PI3K γ . PI3K α and PI3K β are ubiquitously expressed, but PI3K δ and PI3K γ are primarily found in leukocytes.



(1) Wortmannin (2) LY294002

Figure 2. Nonselective PI3K inhibitors used in early biological studies.

Table 1.	Cells	and	Cytokines	Implicated	in	Pathogenesis of
Disease						

disease	main cell types and cytokines implicated in pathogenesis of disease
RA^7	CD4 ⁺ T-cells, macrophages, B-cells, SFs, IFN-γ, TNF-α, IL-6 , IL-12, IL-17, IL-23
SLE ⁸	B-cells autoantibodies, T-cells, neutrophils, macrophages, basophils,
MS ⁹	T-cells, B-cells, macrophages
psoriasis ¹⁰	T-cells (T _h 1), dendritic cells, neutrophils, IL-23, IL-17, IL-22, TNF- α , IFN- γ
COPD ¹¹	CD8 ⁺ T-cells, neutrophils, macrophages, IgG autoantibodies, eosinophils
ALI/ARDS ¹²	neutrophils, T-cells
asthma ¹³	T _h 2 cells, B-cells, mast cells, macrophages, IgE, histamine, IL-4, IL-5, IL-13, TNF-α, IL-1, eosinophils, macrophages, allergen-specific CD4 ⁺ T-cells
IPF^{14}	increases in collagen, fibrosis, TGF- eta
IBD ¹⁵	neutrophils, macrophages, CD4 ⁺ T-cells
MI ¹⁶	leukocyte migration

discovery is to discern which profile will lead to efficacy along with safety appropriate for the particular disease indication.

Table 2. Published IC_{50} (μM) Values for Tool Compounds

	PI3K isoform, IC_{50} (μ M)									
compd	δ	γ	β	α						
1^a	0.009	0.005	0.014	0.001						
2^b	1.05	6.60	0.31	0.73						
3 ^c	0.13	61	16	>200						
4 ^c	0.18	17	11	>200						
5^d	>20	0.03	>20	0.940						
6 ^{<i>a</i>}	0.3	0.008	0.27	0.06						
7^e	0.235	0.83	1.2	1.3						
8 ^a	>20	0.25	>20	4.5						
9^e	1.7	0.07	1.45	0.24						
10 ^c	0.058	0.018	0.35	0.011						
11 ^c	0.048	0.15	0.088	0.008						

^{*a*}ATP concentration equals $K_{\rm m}$.²¹ ^{*b*}ATP concentration equals $K_{\rm m}$.³³ ^{*c*}Tested at 10 μ M ATP.³⁴ ^{*d*}Determined in triplicate.²⁰ ^{*e*}Commercial analysis.³⁵

Highly PI3K-isoform selective compounds are likely to provide the safest bet for becoming drug candidates in conditions that require chronic dosing such as many autoimmune disorders. Although selectivity measurements are an important parameter to consider when developing a selective PI3K small molecule antagonist for chronic inflammatory conditions, physical parameters such as solubility and lipophilicity are variables that have an impact on overall ADME properties and also have to be considered when choosing a compound for further profiling. For example, although the PI3K γ versus PI3K α selectivity of **8** (AS-604850)²¹ was superior to **6** (18 vs 7.5 respectively), compound **6** had a much improved PK profile and was hence chosen for further examination.²¹

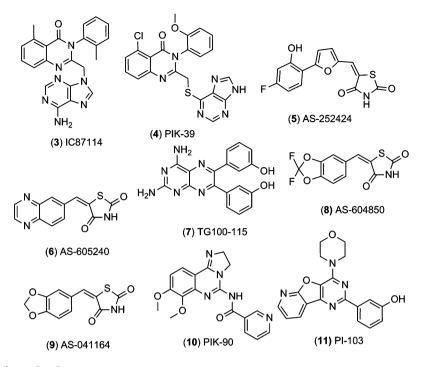


Figure 3. Early inhibitors of PI3K δ and PI3K γ .

The nonredundant roles of PI3K δ and PI3K γ in leukocyte function make them each attractive targets for intervention of immune-mediated diseases. Much has been learned from studies with mice where either or both isoforms were genetically disabled,^{23–26} and knowledge of the KDKI or KO phenotypes informs interpretation of results in in vivo experiments using PI3K small molecule inhibitors.²⁷

The PI3K δ KDKI²⁸ mice show a leukocyte phenotype, with the most pronounced effects seen in B cell function. Thus, in the absence of δ -kinase activity, B-cell proliferation in response to B cell receptor stimulation is inhibited and their function as antigen presenting cells (APC) is impaired. Nearly 50% of the peripheral B-cells and follicular splenic B-cells are eliminated, and there are no MZ or B1 B-cells.²³ Effects observed in the PI3K δ KO mice are similar though less severe, exhibiting impaired lymphocyte activation with developmental defects in B-cells and other cells.

B-CELLS AND PI3Kδ AS TARGETS FOR DISEASE THERAPY

Rituximab (Rituxan) treatment results in the depletion of Bcells, proving efficacious as therapy for non-Hodgkin's lymphoma. More recently, rituximab was also approved for treatment of RA,²⁹ for the first time implicating B-cells as drivers in this disease. The mechanism underlying the success of rituximab's ability to ameliorate autoimmune disease such as RA³⁰ is complex and not completely understood,³¹ but B-cells are involved in cytokine production and the production of autoantibodies.³² Their function as professional APCs in chronic inflammatory conditions may also be underappreciated. While PI3K δ signaling is required for many functions of multiple leukocytes, its strong impact on B cell signaling alone suggests that inhibition of this PI3K isoform may impact the treatment of RA and other autoimmune disorders with B cell components such as lupus. Interestingly, PI3Ky does not appear to have a role in B-cell activation or function.

T-CELLS: PI3K δ AND PI3K γ , TARGETS FOR DISEASE THERAPY

T-Cell function is profoundly affected in PI3K γ and PI3K δ KDKI and KO mice. For example, the differentiation of T_h1 cells, T_h2 cells, and T-cell proliferation to antigen is compromised in PI3K δ KDKI mice, as is IL-10 mediated regulatory T-cell function, although no apparent effect on Tcell development has been noted.²³

PI3K δ plays a nonredundant role in T-cell function, making PI3K δ inhibition an even more attractive target for RA, as well as other autoimmune diseases where T-cells are thought to play a key role in disease pathology.³⁶

Work involving PI3K γ KO mice demonstrated that PI3K γ controls both the survival of thymocytes (progenitors of mature T cells) and the activation of mature T cells.³⁷ There is also a direct role for PI3K γ in eliciting a TCR stimulated response sustaining T-cell and APC interaction.³⁸ Additionally, the migration of T-cells occurs after stimulation of PI3K γ or other class 1A PI3Ks by certain chemokines.^{39,40}

NEUTROPHILS AND OTHER LEUKOCYTES: PI3K δ AND PI3K γ

The role of neutrophils in inflammatory diseases such as RA, COPD, and SLE is well established.²⁸ Neutrophil migration to inflamed tissue and subsequent release of proteases and reactive oxygen species (ROS) cause tissue damage and further activation and recruitment of leukocytes. Although all four PI3K isoforms are expressed in neutrophils, experiments were performed using neutrophils from KO mice and antibodies specific for p110 δ indicating that PI3K δ and PI3K γ are the main players in chemoattractant stimulated neutrophil or macrophage migration. Compound 3 was used to demonstrate that PI3K δ plays an active role in the directional chemotaxis of neutrophil migration by limiting the (direct fMLP induced) formation of PIP3.⁴¹ In vivo studies with 3 and PI3K δ KO mice demonstrated a similar decrease of extravasation of neutrophils

Table 3. PI3K δ , PI3K γ : In Vivo Models of Disease

model	species ^a	modality ^b	PI3K	notes, disease biomarkers	outcome	disease	re
ovalbumin	m	KDKI	δ	IL-4, IL-5, IL-13	decreased	asthma	59
valbumin	m	3	δ	IgE levels, OVA-bound IgE	increased	asthma	59
valbumin	m	3	δ	IgE levels, AHR, IL-4, IL-5, IL-13	decreased	asthma	53
valbumin	m	3	δ	IL-17 expression	decreased	asthma	60
garette-smoke-induced glucocorticoid	m	KDKI	δ	$(\text{GR-}\alpha)/(\text{HDAC-}2)^c$	function restored	COPD	61
-				lung neutrophils	≥50% decreased	asthma	
		КО	γ		no effects		
valbumin ^d	m	7	γ/δ	AHR lung eosinophils	50% decrease 79% decrease	asthma	49
garette-smoke-induced pulmonary ^d	m	7	γ/δ	pulmonary neutrophilia	≥81% decrease	COPD	22, 49
PS-induced pulmonary ^d	m	7	γ/δ	pulmonary neutrophilia	42-95% decrease	COPD	22, 49
OVA-specific pulmonary	m	КО	γ	cell infiltration (neutrophils, eosinophils, lymphocytes, macrophages)	decreased	asthma	46, 47
assive cutaneous anaphylaxis	m	3	δ	PCA response (anti-DNP IgE stimulation) and vascular permeability	decreased	allergy	44
		5	γ	PCA response (anti-DNP IgE stimulation) and vascular permeability	no effect	allergy	44
leomycin pulmonary fibrosis	r	6	γ	BALF leukocyte content	decreased	IPF	62
				collagen content	decreased		
SS colitis	m	KDKI	γ	clinical score vs control	decreased	IBD	63
				IFN- γ , TNF- α	increased		
SS colitis	m	КО	γ	clinical score vs control	decreased	IBD	64
				IL-1 β , TNF- α	decreased		
SS colitis	m	6	γ	clinical score vs control	decreased	IBD	64
				IFN- γ , IL-1 β , TNF- α	decreased		
/BxN serum transfer	m	3	δ	paw diameter, histopath clinical score	strong response	RA	74
		3/KO	δ/γ	paw diameter, histopath clinical score	synergistic response		
		KO/KO	γ/δ	paw diameter, histopath clinical score	synergistic response		
CIA	m	6	γ	clinical score vs control	decreased	RA	21
IA	r	6	γ	clinical score vs control	decreased	RA	21
CII1A	m	6	γ	clinical score vs control	similar to WT	RA	21
IA	m	6	γ	clinical score vs control	40% lower	RA	65
IA	r	3	δ	clinical score vs control	decreased	RA	66
IA	m	INK055 ^e	γ/δ	clinical score vs control	decreased	RA	67
ANTES activation	m	2, 9	γ	neutrophil migration into peritoneal cavity	decreased	RA	33
rrageenan paw edema	m	9	γ	paw swelling	decreased	RA	33
tokine-dependent hTNFtg	m	$KO \times hTNFtg$	γ	synovial mesenchymal fibroblasts	decreased prolif	RA	68
arthritis				MMP3	decreased prolif		
				cartilage damage	decreased		
				subchondral, bone erosion	decreased		
liles assay vascular permeability	r	7	γ/δ	VEGF-induced vascular permeability	inhibited		69
				hindpaw volume	reduced $\geq 62\%$		
Ι	r	7	γ/δ	60 min coronary artery occlusion ^f	≥40% ^g	MI	69
П	р	7	γ/δ	90 min coronary artery occlusion ^f	35% ^g	MI	69
П	m	6	γ	coronary artery ligation and infarct measurement at 14 days	increased infarct size	MI	70
PS induced ALI/ARDS	m	6 , KO	γ	aerosolized LPS; PMN transmigration, vascular permeability	similar effects	ALI/ARDS	12
AE	m	6 , KO	γ	MOG induced; CCL2, CCL5 levels in CNS leukocyte rolling or adhesion	decreased no effect	MS	71
IRL-lpr lupus	m	6	γ	glomerulonephritis	decreased	SLE	72
				: Glucocorticoid receptor- <i>α</i> , HDAC-2: Histone			

Species m: mouse, r: rat, p: pig. Inhibition modality. GR- α : Glucocorticold receptor- α , HDAC-2: Histone deacetylas ^eINK055 (structure not disclosed). ^fReperfusion and infarct measurement at 24 h. ^gPercent reduction in infarct size.

toward sites of inflammation and the prevention of neutrophil adhesion to endothelium cells.⁴² Nonetheless, PI3K γ and

 $PI3K\delta$ do act in concert in the migration, adhesion, and ultimate antimicrobial function of neutrophils. Neutrophils, T-

cells, and B-cells are among many cell types that have been implicated in autoimmune and inflammatory diseases. Other cells that have been thought to play a role, specifically with asthma and atopic disorders, are mast cells and basophils, both of which are activated through their Fc ϵ receptor (FcR ϵ) by IgE bound to allergen. FcR ϵ signaling is dependent on PI3K δ , and its inhibition prevents mast cell and basophil degranulation.^{43,44}

Comparable to PI3K δ , the inhibition of PI3K γ both pharmacologically and genetically had similar inhibitory effects on mast cell degranulation in vitro. For example, comparable inhibitory effect was observed for degranulation of bonemarrow derived mast cells (BMMC) stimulated with Ag/IgE in the presence of inhibitors 3 (PI3K δ selective) and 5 (PI3K γ selective, Figure 3 and Table 2). Degranulation of stimulated mast cells derived from KO or KDKI mice was similarly diminished.^{20,45} PI3K γ and PI3K δ have effects on other leukocytes such as eosinophils. Eosinophil infiltration profoundly reduced in PI3Ky KO animals compared to WT mice in a model of ovalbumin (OVA) specific pulmonary inflammation,^{46,47} while PI3K δ inhibition reduces eosinophil rolling and adhesion to endothelium.48 The dual inhibitor 7 was found to be efficacious in an AHR response OVA-murine asthma model, reducing the level of lung eosinophils by 79%, similar to the effects with dexamethasone.⁴⁹ Other leukocytes have been implicated in inflammatory disorders (Table 1). Indepth discussions of how P3K δ and PI3K γ inhibition may ameliorate activity of these cells have recently been reviewed. $^{50-52}$

TARGETED DISEASES AND EFFECTS OF PI3Kδ AND PI3Kγ INHIBITORS

The role of PI3K δ and PI3K γ signaling in leukocytes is complex. While it is clear that both isoforms play important roles in all leukocytes, the function of each isoform is often associated with signaling through specific cell surface receptors, which gives PI3K δ and PI3K γ distinct and often complementary functions within each cell type.

Several animal models, primarily for rodents, have been used in an attempt to establish a disease association with PI3K δ and PI3Ky signaling whereby kinase activity of one of the two isoforms was either genetically (KO, KDKI) or pharmaceutically (selective compounds) impaired. Several rodent models of RA have been used to show disease amelioration in the absence of either isoform.^{21,33} Similarly, simulating aspects of asthma, acute lung inflammation models using OVA or fMLP as the stimulus were used to show disease modulating effects through inhibition of either PI3K isoform.^{43,53–55} And last, transgenic mice with enhanced class IA PI3K activity have a lymphoproliferative disorder with systemic lupus-like symptoms⁵⁶ while PI3K γ selective inhibitor 6 showed efficacy in a spontaneous mouse model of lupus. Many other models of autoimmune and inflammatory diseases have been tried, and disease modifying effects have been shown by inhibiting either PI3K isoform. Some of these are summarized in Table 3. Even though there are many preclinical data showing the impact of PI3K δ or PI3K γ in rodent models of human disease, we have to bear in mind that rodents are in every aspect quite distinct from humans. In addition, few of these rodent models are spontaneous and chronic in nature but are instead induced by administration of specific antigens leading to acute inflammation. While these models may reflect certain aspects of human disease, they do not accurately reflect the complexity

and heterogeneity associated with human disease pathology. So despite the intriguing biology associated with both PI3K isoforms that holds promise for inflammatory disorders and despite the multitude of preclinical models that seem to substantiate that promise, we will have to await the outcome of clinical trials to determine whether selective inhibitors of PI3K δ or PI3K γ are indeed safe and efficacious.

SAFETY CONCERNS FOR PI3Kγ AND PI3Kδ AS A TARGET FOR INFLAMMATION AND AUTOIMMUNE DISEASE

Death of the PI3Ky KO mice was observed when high doses of vaccinia virus (VV) were employed in a model examining the migratory ability of CD8 T-cells. In contrast, WT mice had little trouble clearing the infectious agents at the same viral loads. Infection is a concern for PI3Ky inhibitors in any indication.⁵⁷ There are also potential concerns with targeting PI3K δ , as the PI3K δ deficient mice develop a mild, subclinical IBD/colitis phenotype possibly due to suppression of IL-10 in these mice.²³ Whether this phenotype would be seen with a selective compound and whether it will translate into humans are currently not known. There is evidence in PI3K δ KDKI mice that points to suppression of some T_{reg} cell functions; this could impact autoimmunity.⁵⁸ There are several publications suggesting that PI3K δ inhibition will lead to an increase in IgE, which may be counterindicative to using these inhibitors for the treatment of allergic diseases.⁵⁹ However, there does seem to be conflicting evidence for this outcome⁵³ (Table 3).

DUAL INHIBITION OF PI3K δ and Pi3K γ

Although PI3K γ and PI3K δ inhibitors have shown promise in preclinical disease models and suggest that a dual inhibitor may have synergistic affects, it remains unclear whether a dual inhibitor would be safe. Mice in which both isoforms are inactive have profound reductions in thymus size, and total thymus cell count. Their peripheral T cell compartments are almost completely absent,⁷³ while mice with individual KOs of p110 γ and p110 δ have relatively normal thymi and thymocytes. In addition, crossing the p110 γ KO with a p110 δ KDKI²⁶ leads to not only severe lymphopenia but also homeostatic peripheral expansion of Th2 cells, with subsequent type 2 inflammation in various organs. This is characterized by increases in IgE secretion in serum and eosinophilic infiltration into the mucosal organs leading to pronounced inflammation. However, although the PI3K γ/δ deficient mice developed homeostatic peripheral expansion of Th2 cells, dual pharmacological inhibition did not increase the Th2 response but decreased the T_h1 cytokine release. It is possible therefore that the observations in genetically dual-deficient mice could result from developmental defects that would not be recapitulated in mature animals under pharmacological intervention. To substantiate that notion, in a murine inflammatory arthritis model using PI3K γ KO mice dosed with 3 a pronounced synergistic effect on disease amelioration was shown.⁷⁴

INHIBITORS OF PI3K γ AND PI3K δ

A great deal of effort in the past several years has been directed at discovering selective inhibitors of PI3K δ and PI3K γ . Many institutions have entered the fray, and the field has expanded rapidly. Additionally, dual inhibitors have also become an attractive target for some.⁷⁵

Table 4. Potency	and Isoform	Selectivity:	Examples	of PI3K δ	Inhibitors	from	the Literature ^{<i>a</i>}
------------------	-------------	--------------	----------	------------------	------------	------	------------------------------------

			bioc	hemical select	tivity			
compd	organization	biochemical PI3Kδ, IC ₅₀ (μM)	γ/δ	$lpha/\delta$	β/δ	cellular PI3Kδ, IC ₅₀ (μM)	refs and notes	
3	Gilead ¹⁵²	0.5	58	>200	150		200 µM ATP	
12	Roche	0.003	25	1	11			
13	Roche	<0.1	>20					
14	Roche	$0.0049 \ (K_{\rm i})$		101		0.017	cell: HWB CD69; WO2006046035	
15	Roche	$0.00046 \ (K_{\rm i})$		256		0.003	cell: HWB CD69; WO2012007493	
16	Roche	$0.000576~(K_{\rm i})$		175		0.0052	cell: HWB CD69; WO2012007493	
17	Roche	0.00343		>350		0.041	cell: HWB CD69; WO2011101429	
18	Roche	0.00274		64		0.0178	cell: HWB CD69; WO2010138589	
19	Novartis	0.005	10-40	10-40	10-40	0.015	WO2008000421	
20	Lilly	0.040	187	1447	126		S-isomer preferred	
21	Lilly	0.006	28	528	123		S-isomer preferred	
22	Lilly		≥10	≥10	≥10	0.665	cell: neutrophil (PMN) elastase release	
23	Lilly		≥10	≥10	≥10	0.665	as above (typical example)	
24	Amgen					0.002	cell: B-cell proliferation; WO08118454	
25	Amgen					0.0005	cell: B-cell proliferation; WO09118468	
26	Amgen	$0.00285~(K_{\rm i})$					WO2010151791	
27	Amgen	$0.00105 (K_i)$					WO2010151737	
28	Amgen	$0.00226 (K_i)$					WO2010151737	
29	Amgen	$0.005 (K_i)$					WO2010151735	
30	Amgen	0.1059					US 7705018, WO2008118455	
31	Amgen	$0.0002 (K_i)$					WO2011123751	
37	Takeda	<0.1	≥ 1				(typical example) WO09046448	
38	Takeda	<0.1	≥ 1	≥10	≥100		potent in a rat CIA model 60 mg/kg b.i.d.	
39	Takeda	<0.1	>100	>100	>100	<0.1	Cell: B-cell proliferation; WO2010129816	
40	Takeda	<0.050		≥20	≥20		WO2010006086, potent against m-TOR	
41	Takeda	0.040	2.8	1.3	16.6		ref 111	
42	Takeda	0.001	185	788	23		ref 111WO2010036380	
43	Takeda	0.0005	8	274	18		ref 111, WO2010036380	
44	Takeda	0.0003	24.7	3643	1157	0.500	cell: (EC ₅₀) proliferation assay ¹⁰⁹	
45	UCSF	0.010	16	1000	49		ref 111	
46	UCSF	0.0007	47	1857	314		ref 111	
47	UCSF	0.007	186	12000	107		ref 111	
52	AUL^{b}	>0.1		1.0-10	≥10		WO2009120094	
^{<i>a</i>} For str	ructures, see F	igures 3–6. ^b Auckland Un	iservices Lte	d.				

It is difficult to discern what constitutes a selective inhibitor. In general, selectivities from 10- to 50-fold seem a reasonable starting point, but some examples may fall below this threshold. The categorization is complicated by the fact that precise IC_{50} values are lacking from the many published patent applications. Refer to Tables 4–7 for the summary of data (references) for structures found in Figures 4–9.

PI3K\delta SELECTIVE INHIBITORS

Roche/Genentech/Piramed has been active in developing PI3K inhibitors, although most patent applications disclose compounds specifically targeting PI3K α . Compound **12** (GDC-0941)⁷⁶ (Figure 4 and Table 4), which entered phase II clinical trials for the treatment of metastatic breast cancer and non-small-cell lung cancer,⁷⁷ is a prominent example.⁷⁸ Several patent applications showcase a series of structurally related thienopyrimidines as PI3K δ selective inhibitors such as **13** (Figure 5).^{79,80} Some of these compounds are highly soluble (40–100 μ M) with low hepatocyte clearance, reasonable permeability, and low CYP-450 inhibition and induction activity. They are potent in a biochemical assay (PI3K δ , IC₅₀ < 0.1 μ M) with generally <90% plasma protein binding and selectivity of >20-fold over PI3K γ in some cases. Morpholino-

thienopyrimidine 14^{81} also has low micromolar activity in a PI3K δ human whole blood (HWB) assay. These compounds have efficacy in a CIA model, although no detail is provided.⁸² Additional compounds from Genentech/Roche, benzoimida-zolpurine 15,⁸³ indolpurine 16,⁸³ and indolpyridopyrimidine 17⁸⁴ are very potent in enzyme and HWB assays. Morpholinofuropyrimidine 18⁸⁵ is one of the more potent analogues described in an application exemplified by 11 analogues having IC₅₀ values in a HWB assay of 0.0178–0.0659 μ M. Additionally, selectivities over PI3K α in an enzyme assay are reported, from 14- to >300-fold.

Other PI3K δ selective inhibitors have been disclosed by Novartis, Lilly, Amgen, Takeda (Intellikine) UCSF, UCB Pharma, Auckland Uniservices, Zenyaku Kogyo, Xcovery (Tyrogenex), Chemical Diversity Research Institute, Serono Astellas Pharma, Karus Therapeutics, GlaxoSmithKline, Incyte, Incozen/Rhizen, Infinity, Exelixis, Respirvert, and Gilead (formerly Calistoga, formerly ICOS). Novartis has claimed a group of compounds exemplified by thiazolylurea **19**.⁸⁶ This compound is potent in a scintillation assay of PI3K δ activity (IC₅₀ = 0.005 μ M) with a selectivity profile over the other isoforms of 10- to 40-fold. Additionally, **19** has a potency of 0.015 μ M in a B-cell proliferation assay. Lilly described

Table 5. Potency and Isoform Selectivity: Examples of PI3K δ Inhibitors from the Literature^a

			bioche	mical sel	ectivity		
compd	organization	biochemical PI3Kδ, IC ₅₀ (μM)	γ/δ	α/δ	β/δ	cellular PI3K <i>δ</i> , IC ₅₀ (μM)	refs and notes
53	Zenyaku Kogyo	0.0046	10.6	3.5	9.5		WO2002088112
54	Xcovery	<0.03	>0.1	>0.1	>0.1		WO2010005558
55	$CDRI^{b}$	2.1	33.8	6.0	38		WO2009011617
56	EMD Serono	0.027				0.016	cell: pAKT of B-cells; WO2007023186
57	EMD Serono	0.090	111	4.4	111		ref 111, WO2007023186
58	EMD Serono	0.034	588	101	124		ref 111
59	Astellas Pharma	0.0036		444		0.00062	cell: anti-IgM B-cell thymidine incorporation
61	Karus	0.26	>38	>38	13		WO2011135351
62	GlaxoSmithKline	pIC ₅₀ > 7.0	>10	>10	>10		WO2010125082
64	GlaxoSmithKline	$pIC_{50} > 7.0$	>10	>10	>10		WO2010125082
65	Incozen/Rhizen	<0.25	d	d	d		
66	Respivert	0.004	72	492		0.57	cell: H ₂ O ₂ induced pAKT; WO 2011048111
67	Incyte	<0.05	>10	>10	>10	< 0.05	cell: B-cell proliferation; WO2011008487
68	Incyte	<0.2				<0.5	cell: B-cell proliferation; WO2011075643
69	Incyte	<0.1				<0.1	cell: B-cell proliferation; WO2011130342
70	Incyte	<0.05					US20110312979
71	Incyte	<0.05				<0.05	cell: B-cell proliferation; WO2011075630
XL499 ^c	Exelixis	0.0075	190	96	111	0.066	cell: inhibition of pAKT(T308) (Raji cells)
72	Exelixis	<0.05					WO2012037226
73	Exelixis	<0.05					WO2012037204
74	Gilead	0.009	21	248	49	0.0061	cell: (EC ₅₀) human B lymphocyte
75	Gilead	$0.007 \ (K_{\rm i})$	61	2963	119		WO2011011550
^{<i>a</i>} For struc	ctures, see Figure 7.	^b Chemical Diversity Resea	rch Inst	itute. ^c St	tructure	not disclosed. ^d <25% i	nhibition at 1 μM

Table 6. Potency and Isoform Selectivity: Examples of PI3K γ Inhibitors from the Literature^a

			hiach	emical sele	ctivity	
		1. 1 1				
compd	organization	biochemical PI3Kγ, IC ₅₀ (μM)	δ/γ	$lpha/\gamma$	β/γ	refs and notes
76	Chemical Diversity Research Institute	0.0059		6339	5915	WO2009011617
77	Serono	0.013				US20080188531
78	Pfizer	0.054	6.3	0.4	20.4	$10 \ \mu M \ ATP^{34}$
		0.001				20 μM ATP, WO2004052373
79	Pfizer	0.003				20 μM ATP, WO2004108713
80	Pfizer	0.197				Streptococcal cell-wall-induced rat paw edema model $ID_{50} = 16.3$ mg/kg (30 mg/kg 52% inhibition)
81	Cellzome	<0.1	>10	>10	>100	WO2010092015, WO2008025821
82	Vertex	$0.1 (K_i)$				WO2008027584
83	Vertex	$0.003 \ (K_{\rm i})$				WO2010096389
84	Vertex	$0.01 \ (K_{\rm i})$		1-10		WO2009129211
85	Vertex	$0.003 (K_i)$		<33		WO2010135014
86	Novartis	<2.0				WO2007134828
87	Novartis	0.016	40	189		WO2005021519, US2008280871
88	Novartis	0.012				WO2009115517
89	Bayer	<0.1			1-5	WO2004029055
90	Chroma Therapeutics	<0.1				WO2007129048
91	Boehringer Ingelheim	<0.6				US20090093474
92	Applied Research Systems	0.006				cell: pAKT macrophages, ELISA; <0.010 μM, WO2006024666
94	Shionogi	0.027				WO2010027002
95	Exelixis	0.018	24.2	38.3	114.4	cell IC ₅₀ = 0.773 μM^{170}
96	Exelixis	0.034	10.1	12.2	33.8	cell IC ₅₀ = 0.805 μM^{170}
^{<i>a</i>} For str	ructures, see Figure 8.					

thienopyrimidinones **20**, **21**,⁸⁷ pyridopyrimidinone **22**, and pyrazolopyrimidinone **23**,⁸⁸ similar in structure and isoform selectivity to **3**. Amgen has compounds containing a central core quinoxaline 24^{89} or quinoline 25^{90} and a purine or pyrazolopyrimidine side chain. No selectivity data are given

though the compounds listed have cellular activity down to 0.0005 μ M. Compounds with biochemical potency against PI3K δ (K_i) in the single digit nanomolar range are exemplified by quinolines **26**,⁹¹ **27**,⁹² **28**,⁹³ and pyridopyrimidinone **29**.⁹⁴ Compounds **28** and **29** contain an unusual aminopyrimidine

Table 7. Potency and Isoform	Selectivity F	Examples of PI3K δ/γ	Inhibitors from	the Literature ^a
Tuble 7. Totelley und 15010111	beleeuvity. I	Dampies of 11510/7	minutions nom	the Diteruture

compd	organization	biochemical PI3Kδ, IC ₅₀ (μM)	γ/δ	$lpha/\delta$	β/δ	cellular PI3Kδ, IC ₅₀ (μM)	refs and notes
7	TargeGen	0.235	3.5	5.5	5.1		
97	UCB Group	0.032	2.4	9.2	16.9	0.111	cell: neutrophil inhibition superoxide
98	UCB Group	0.139	0.77	5.3	9.5		
99	UCB Group	0.014	3.7				
100	Millenium	<0.1 ^b					WO2009154741
101	Millenium	< 0.1 ^b					WO2009154741
102	GlaxoSmithKline	$pIC_{50} \ge 5^b$				$pIC_{50} \ge 5^c$	cell: human PBMC
103	GlaxoSmithKline	$pIC_{50} \ge 5^b$				$\text{pIC}_{50} \ge 5^d$	cell: pAKT T-cells; WO2009147190
104	GlaxoSmithKline	$pIC_{50} \ge 5^b$				$\mathrm{pIC}_{50} \ge 6^d$	cell: pAKT T-cells; WO2009147189
105	GlaxoSmithKline	$pIC_{50} \ge 5^b$					WO200912958
106	GlaxoSmithKline	$pIC_{50} \ge 5^b$				$\text{pIC}_{50} \ge 6^d$	cell: pAKT T cells; WO2009147188
107	GlaxoSmithKline	$pIC_{50} \ge 5^b$					WO2011067365
108	GlaxoSmithKline	$pIC_{50} \ge 5^b$					WO2011067366
109	UCSF	0.009	2.2	1000	77		ref 111
110	Takeda	<0.1	$IC_{50} < 0.1$	>10	>10	<0.1	WO2009088990
INK055 ^e	Takeda	0.005	1.8	>2000	>2000		refs 67, 109
^{<i>a</i>} For struct	ures, see Figures 3	and 9. b PI3K δ and/or PI3	Kγ. ^c Presume	ed PI3Kδ.	^d Presume	ed PI3Kγ. ^e Structure n	ot disclosed.

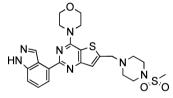


Figure 4. 12 (GDC-0941).

side chain. Amgen has also published patent applications on many other novel structures, some with a pyrimidine instead of a purine but all with diverse core moieties. These are exemplified by quinoline 30,⁹⁵ benzimidazole 31,⁹⁶ thienopyr-idine 32,⁹⁷ 1,5-naphthyridine-4-carboximide 33,⁹⁸ and quino-lines 34,⁹⁹ 35,¹⁰⁰ and 36,¹⁰¹ although no biological data were provided. Amgen has advanced one PI3K δ inhibitor AMG-319 (structure not disclosed) into phase I clinical trials for lymphoid malignancies.¹⁰² Intellikine (now Takeda) specialized in PI3K and m-TOR inhibition and indicated that they have selective PI3K δ inhibitors.¹⁰³ The relevant structures were not disclosed, but they have described various substituted quinazolines such as 2-methylaminoquinazoline 37.¹⁰⁴ Selectivity for PI3K δ and PI3Ky relative to the other isoforms and also m-TOR is claimed, although no specific biological data are provided. Methylisoquinolinone 38^{105} is the most studied analogue in an application that deals with a series of isoquinolinones, although **38** could be considered a dual PI3K δ /PI3K γ inhibitor. There is some interesting biological data, including a CIA model with a pronounced reduction in mouse ankle diameter and histopathological score at the 60 mg/kg dose. Other examples in the applications have a greater level of selectivity favoring PI3K δ . Another application describes isoquinolines such as 39^{106} with potencies of <100 nM against PI3K δ and selectivities over the other isoforms of >100-fold in an enzyme assay. A series of compounds similar in structure to 38 but with a small group such as an isopropyl or cyclopentane replacing the isoquinolinone functionality have also been claimed by

Takeda (Intellikine). These are exemplified by pyrazolopyrimidine 40.¹⁰⁷ Although 40 is selective against PI3K α and PI3K β , it is also potent toward mTOR (<0.05 μ M). Compounds of this type were reported to be active in a lupus model, perhaps indicating that they are not all that selective over $PI3K_{\gamma}^{1.08}$ An example with a more extensive profile is the isoquinolinone 44 (INK007).¹⁰⁹ This compound was found to be potent and selective and inhibited the growth of PDGF induced FLS (fibroblast-like synoviocytes). Shokat and co-workers at UCSF have revealed a series of selective PI3K δ inhibitors, exemplified by quinazolinones 45,¹¹⁰ 46 (SW-13),¹¹⁰ and 47 (SW-30),¹¹⁰ that directly target the PI3K affinity pocket, enhancing potency considerably.111 UCB Pharma has described a series of quinolines, as PI3K δ inhibitors exemplified by 48,¹¹² 49,¹¹³ 50,¹¹⁴ and 51,¹¹⁵ but no biological data were provided. Auckland Uniservices Ltd., working to develop PI3K α inhibitors containing triazine and pyrimidine, also discovered several compounds that have reasonable selectivity toward PI3K δ , as exemplified by the morpholinotriazine **52**.¹¹⁶ Zenyaku Kogyo has been developing compound 53 (ZSTK474)¹¹⁷ for oncology that has potent antitumor activity in vivo.¹¹⁸ It has some selectivity toward the PI3K δ isoform.¹ Compound 53 also demonstrated efficacy in a CIA model of RA, both in a semitherapeutic and therapeutic setting.¹²⁰ Xcovery has disclosed compounds with selectivity toward PI3K δ . One of their most potent and selective compounds is the morpholinopurine 54.¹²¹ The Chemical Diversity Research Institute revealed substituted pyrimidinones, including pan inhibitors as judged by the selectivity data. There are examples, however, that exhibit strong selectivity toward PI3K δ or PI3K γ exemplified by furopyrimidinones 55 and 76¹²² (Figures 7 and 8, respectively). This is an interesting example of a small change in structure having a large impact on the selectivity characteristics, the difference being a 4-OMe-phenyl group versus an unsubstituted benzyl group. The changes in 76 increased potency toward PI3K γ and increased selectivity over the PI3K α and PI3K β isoforms. Serono has been active in discovering

Perspective

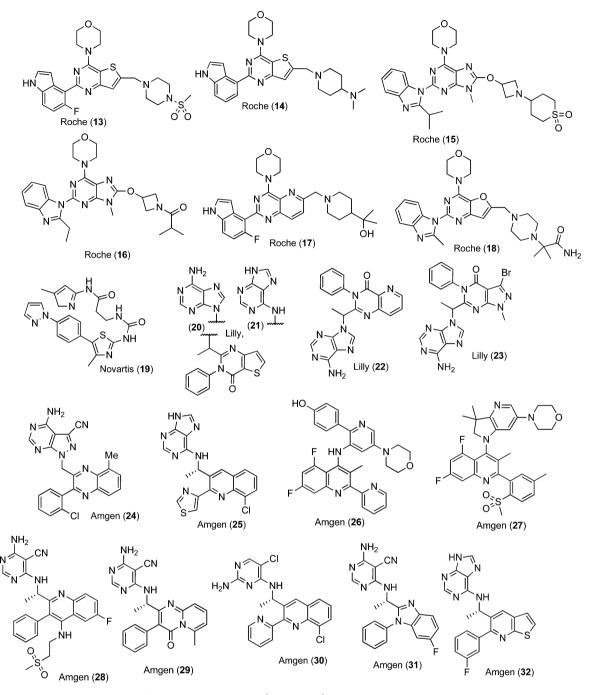


Figure 5. Examples of PI3K δ inhibitors from the recent literature (see Table 4).

compounds that inhibit PI3K δ and PI3K γ . Quinoxaline 56¹²³ is a typical example, which is potent in a biochemical assay and also active in an anti-IgM stimulated peripheral blood mononuclear cell (PBMC) cellular assay. Several compounds including 41 (DL06),¹¹¹ 42 (INK654),¹²⁴ 43 (INK666),¹²⁴ 46, 47, 57 (AS5),¹²³ and 58 (AS15)¹¹¹ have been extensively profiled including multiple PI3K δ cocrystal structures from researchers at the University of Cambridge, UCSF, Takeda (Intellikine), and Merck-Serono.¹¹¹ Quinoxaline 57 has a high level of PI3K δ selectivity over the PI3K γ and PI3K β isoforms. In addition these workers also studied 42 a quinoline analogue that has PI3K δ selectivity relative to PI3K γ and PI3K α , less so to PI3K β . Tetrahydroquinazolinone 58 has an unusual fivebond linkage from the core moiety to a pendent quinoxaline (see next section). Astellas Pharma has examples of potent PI3K δ inhibitors. One of their most potent is morpholinotriazine **59**¹²⁵ with an IC₅₀ in B-cells of <1 nM. Karus has disclosed compounds such as morpholinobenzoxazinone **60**¹²⁶ and indoylisoquinoline **61**¹²⁷ that may be PI3K δ and/or dual PI3K δ /PI3K β inhibitors. No biological data were provided for **60**, but **61** is 13-fold selective over PI3K β and >38-fold over PI3K γ and PI3K α . Karus has revealed that they are pursuing two inhibitors KAR4139 (structure not disclosed), a dual PI3K δ /PI3K β inhibitor, and KAR4141(structure not disclosed), a selective PI3K δ inhibitor for cancer/inflammation and inflammation, respectively.¹²⁸ GlaxoSmithKline has disclosed compounds¹²⁹ such as indoylindazole **62**¹³⁰ and furo[3,2-*b*]pyridineindazole **63**¹³¹ that are potent toward

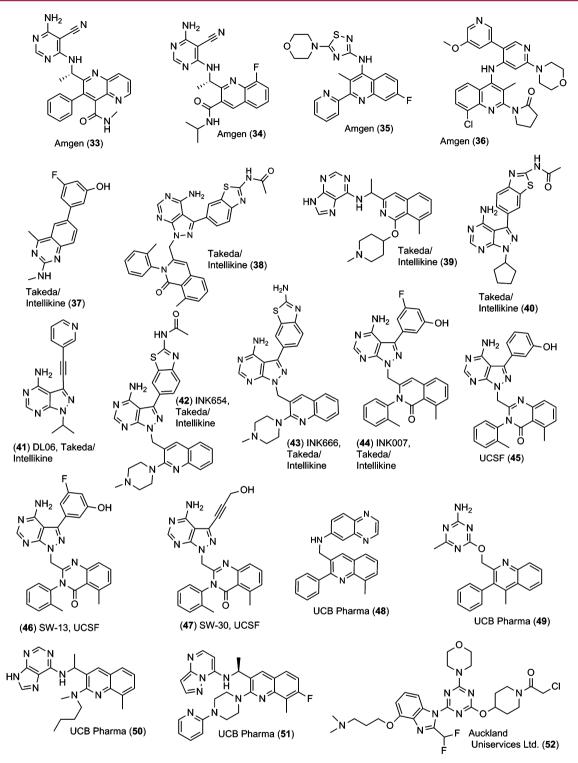


Figure 6. Examples of PI3K δ inhibitors from the recent literature (see Table 4).

PI3Kδ. Compound **62** is at least 10-fold selective over the other isoforms. Additionally, they have recently disclosed various salt forms and polymorphs of oxazoloindazole **64**,^{130,132} another inhibitor with greater than 10-fold PI3Kδ selectivity. Interestingly, GlaxoSmithKline has started a phase I clinical trial targeting asthma with GSK2269557 (structure not disclosed).¹³³ Incozen/Rhizen has described a patent that showcases a series of 2,3-substituted 4*H*-chromen-4-ones such as **65**¹³⁴ with potency toward the PI3Kδ isoform. Respirvert disclosed quinoxoline compounds such as **66**,¹³⁵ which is potent against PI3K δ and selective over PI3K γ and PI3K α . Incyte has also been active in discovering PI3K δ inhibitors. They describe at least five compounds, which have potency in a B-cell proliferation assay of less than 50 nM. These compounds, including the thiazolopyrimidinone **67**,¹³⁶ are potent on an enzyme assay with at least 10-fold selectivity over all the other isoforms. Compound **67** was potent (<50 nM) in a proliferation assay using a diffuse large B-cell lymphoma cell line. Incyte has additional patent applications with a six–five fused core, including both imidazo[1,2-a]pyridine **68**¹³⁷ and

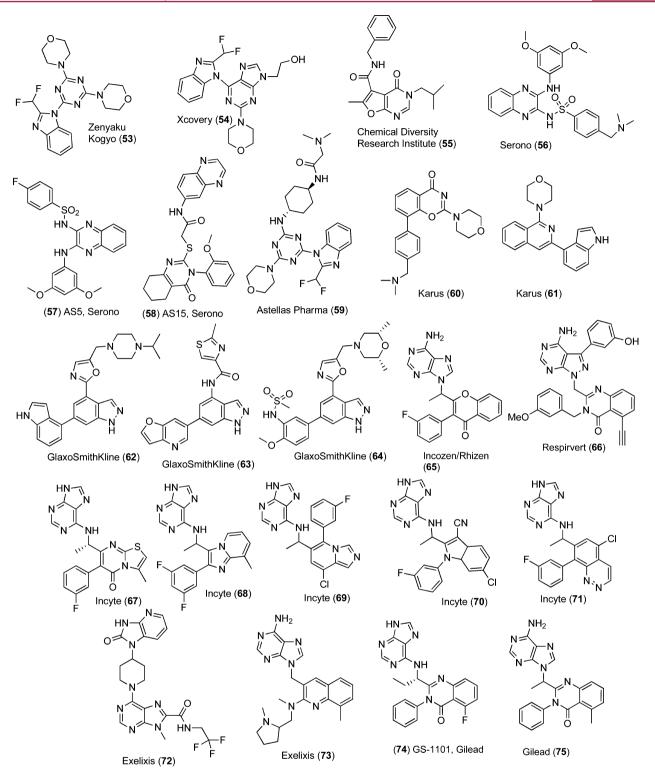


Figure 7. Examples of PI3K δ inhibitors from the recent literature (see Table 5).

imidazo[1,4-*a*]pyridine **69**.¹³⁸ These were found to be potent in a B-cell proliferation assay at <500 and <50 nM, respectively. Additional patent applications from Incyte have exemplified indolylpurine **70** and cinnolinylpurine **71**.¹³⁹ The key feature in these applications remains the pendent ethylaminopurine. Pathway Therapeutics claims to have an inhibitor that is potent against PI3K δ and >100-fold selective.¹⁴⁰ No structures are disclosed, but their previous patent literature indicates compounds that have pan-inhibitor character reminiscent of

53 and **54**. Exelixis has partnered with Merck in the development of XL499 (structure not disclosed), a selective PI3K δ inhibitor targeting hematological malignancies and inflammation. Although the structure has not been disclosed, data surrounding this compound have been released.¹⁴¹ The compound demonstrated high selectivity and reasonable PK, *F* of 48–99.9% in animals (cynomolgus monkey, rat, mouse, dog). It was potent in cell assays and displayed effectiveness in a passive cutaneous anaphylaxis mouse model. The compound

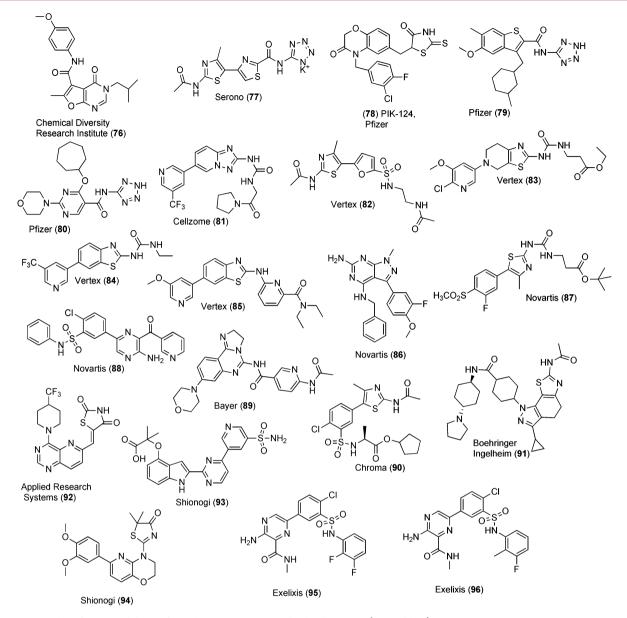


Figure 8. Examples of PI3K γ inhibitors from the recent patent and other literature (see Table 6).

appeared well tolerated in a 14-day rat toxicology study with no clinical signs or significant change in animal weight. Exelixis has described specific PI3K δ inhibitors in two patent applications. The first lists a series of 6,9-disubstituted purines that generally incorporate a 6-piperidinylimidazopyridine such as 72.¹⁴² Of the 397 compounds described, 146 inhibit PI3K δ at <50 nM. The other patent application describes 191 analogues of which 40 have potency toward the PI3K δ isoform of less than 50 nM. Quinoline 73¹⁴³ is such an example.

The most advanced group remains Gilead (formerly Calistoga) with two compounds in clinical trials. Quinazolinone 74 (GS-1101 formerly CAL-101)¹⁴⁴ is a PI3K δ selective compound that has been in several phase I/II trials for hematologic malignancies, including combination therapy with rituximab.^{145,146} Promising results have been disclosed. In a phase I study involving 57 patients with hematological malignancies (NHL, CLL, AML) significant clinical response rates were seen at doses of 50, 100, 200, and 350 mg b.i.d., although there was variability among the types of malignancies. For example, although there was no improvement in the AML

cohort, response rates of 56% was found in the NHL cohort. The dose limiting toxicity was a reversible increase of serum transaminases.¹⁴⁷ More clinical data relating to 74 are emerging, but some aspects of the compound itself require elaboration. The compound was found to be well tolerated at 400 mg q.d. and 200 mg b.i.d. for up to 7 days of dosing in healthy volunteers. In patients with lymphoid malignancies dose levels of 350 mg were tolerated for several months. An MTD was not reached, but exposures were not dose proportional at these levels. There were minimal gains in plasma exposure beyond a dose of 150 mg b.i.d. Steady state trough levels from b.i.d. dosing of \geq 150 mg could be maintained at >10-fold over the HWB EC₅₀ of 62 nM.¹⁴⁸ The PI3K δ inhibitor 74 has entered phase II trials for CLL in combination therapy with rituximab. It was also examined in a phase I trial of allergic rhinitis involving doses of 100 mg b.i.d. for 7 days.¹⁴⁹ Another compound CAL-263 (structure not disclosed) presumed to be an analogue of 74 seems to be intended primarily for inflammatory diseases. This drug candidate was also entered in a similar allergic rhinitis phase I

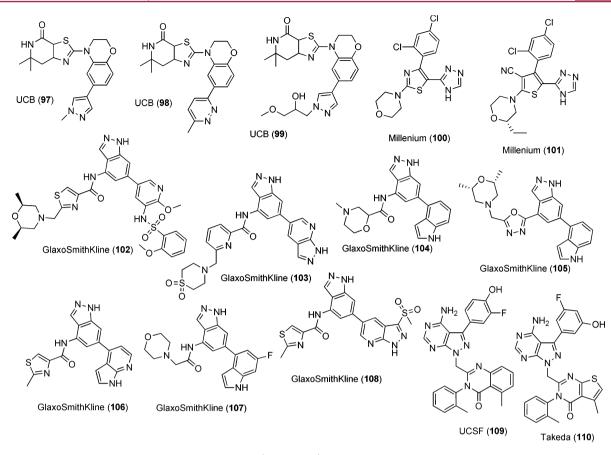


Figure 9. Examples of PI3K δ/γ inhibitors from the literature (see Table 7).

trial and at >10-fold lower dose (10 mg q.d.).¹⁵⁰ Gilead (Calistoga) has recently published a patent application exemplified by the potent and isoform selective quinazolinone 75^{151} for use in certain liver disorders.

ΡΙ3Κγ SELECTIVE INHIBITORS

Selective inhibitors of PI3K γ are also of great interest. In addition to the Serono compounds 5, 6, 8, and 9 (AS-041164), Serono has also published on bis-thiazoles. Tetrazole 77¹⁵³ is one of their potent examples. Pfizer/Warner Lambert has several published patent applications, with benzoxazinone 78^{154} and benzothiophene 79^{155} as representative examples. The selectivity over PI3K α may be an issue, as 78 indicates (α/γ = 0.4, Table 6). However, pyrimidine 80¹⁵⁶ demonstrated reasonable activity in a Streptococcal cell wall (SCW) paw edema model of arthritis and had activity in a CIA model at 30 mg/kg (52% reduction in paw edema). Cellzome has described compounds that have potency against PI3K γ (<0.1 μ M) and various selectivity over the other isoforms, with some examples of dual PI3K γ/δ inhibitors. Many of these compounds have selectivity over the PIKK DNA-PK. Their most potent and selective analogue reported is triazolopyridine **81**.¹⁵⁷ Vertex has examples of potentially PI3K γ selective inhibitors such as furylthiazole **82**,¹⁵⁸ tetrahydrothiazolopyridine **83**,¹⁵⁹ benzo-thioazolyl-urea **84**,¹⁶⁰ and benzothiazolylamine **85**.¹⁶¹ Novartis has patent submissions on PI3K γ since 2003. The diaminopyr-azolopyrimidine **86**¹⁶² had some potency toward the PI3K γ isoform, but thiazole **87**¹⁶³ had an IC₅₀ of 16 nM and was 39fold selective over PI3K δ and 189-fold selective over PI3K α , although no selectivity data were provided against PI3K β . In 2009 they disclosed a series of substituted pyridines or

pyrazines such as aminopyrazine 88¹⁶⁴ that appear quite potent against PI3Ky. Bayer, Chroma Therapeutics, and Boehringer Ingelheim have programs aimed at discovering PI3Ky inhibitors. Although representative compounds dihydroimidazoquinazoline **89**,⁵⁴ thiazole **90**,¹⁶⁵ and dihydrothiazoloindazole 91¹⁶⁶ may be quite potent, no selectivity data have been published. Applied Research Systems has described a series of pyridinemethyleneazolidinones. A representative example is 92^{167} with single digit nanomolar potency in a biochemical assay against PI3K γ and IC₅₀ < 10 nM in a p-AKT ELISA cell assay. Shionogi has been active in developing PI3K γ inhibitors. The pyrimidine 93¹⁶⁸ is potent in a whole cell assay with \geq 95% inhibition of AKT phosphorylation at 10 μ M. The compound showed reasonable physical chemical properties as well; however, no selectivity data were provided. They have also claimed a series of pyridooxazines such as 94^{169} with potency in a biochemical assay in the double digit nanomolar range. Exelixis has published a series of analogues exemplified by Nmethylpyrazine-2-carboxamides 95 and 96.¹⁷⁰ These compounds, while exhibiting considerable selectivity over the other isoforms, had excellent physicochemical properties. The high oral bioavailability exhibited by 95 (76.7%) and 96 (66.2%) in rats was attributed to the high solubility and permeability of the analogues. Both compounds also demonstrated effectiveness in a mast cell degranulation in vivo mouse model.

DUAL PI3K γ AND PI3K δ INHIBITORS

Though it is occasionally difficult to discern from the patent literature whether a program is targeting selective PI3K δ , PI3K γ , or dual inhibition, there are several examples where a dual inhibitor is being specifically pursued. TargeGen³⁵

produced 7, a dual inhibitor of PI3K γ and PI3K δ in phase I/II clinical trials against ischemic reperfusion injury following myocardial infarction. As mentioned previously, compound 7 was also studied for use in asthma and COPD, where it was effective in a cigarette-smoke-induced pulmonary mouse model. In the course of 3 days over several dosing regimens, it was found that neutrophil accumulation in BALF was decreased significantly relative to smoke treatment alone. Another in vivo experiment found that induction by LPS for enhancement of pulmonary neutrophilia could be ameliorated by aerosolized 7. In this model decreases in BALF neutrophils of 42% were found compared to dexamethasone (70% reduction).⁴⁹

The UCB group has reported a few compounds with noted selectivity for PI3K δ over the other isoforms, although generally these compounds had similar potency between PI3K γ and PI3K δ .¹⁷¹ For example, tetrahydrothiazolopyridinone 97^{172} has a PI3K δ IC₅₀ of 32 nM but is only 2.4-fold less potent toward PI3Ky. The compound has reasonable PK, as demonstrated in Han–Wistar rats (Cl = 7 mL min⁻¹ kg⁻¹, F =97%). The structurally related 98 displayed similar selectivity and PK in Han–Wistar rats (Cl = 13 mL min⁻¹ kg⁻¹, F = 66%). However, both these compounds have poor solubility and were less than 10-fold selective over PI3K α . Incorporation of a solubilizing group providing 99 increased the solubility and decreased the plasma protein binding. Inhibitor 99 has an ED_{50} of 5 mg/kg in a rat in vivo model of CD3 induced IL-2 ³ Wilex has partnered with UCB to develop WX-037 release.1 (structure not disclosed), a dual PI3K δ/α inhibitor for solid tumors. Millennium has disclosed thiazole 100 and thiophene 101 for targeting inflammation.¹⁷⁴ Few biological data were disclosed, other than a range of IC_{50} values against the isoforms. GlaxoSmithKline has disclosed a series of carboxamide substituted indazoles claimed to be useful for COPD, asthma, and other inflammatory diseases. Several compounds such as pyridylindazole 102¹²⁹ are potent in an enzyme assay against various PI3K isoforms and against PI3K δ in a human PBMC cell assay with a mean pIC₅₀ of 5 or greater. Similar compounds, for example, picolinamide **103**,¹⁷⁵ indazole **104**,¹⁷⁶ oxadiazole **105**,¹⁷⁷ thiazolecarboxamide **106**,¹⁷⁸ morpholinoacetamide **107**,¹⁷⁹ and pyrozolopyridine **108**,¹⁸⁰ may have activity against PI3K γ and PI3K δ in an enzyme assay or a T-cell assay (presumably PI3K γ) with a pIC₅₀ of 5–6 or greater. Shokat and co-workers discovered an interesting selectivity differential. A few minor structural changes with a phenol moiety in the affinity pocket caused a shift in the PI3K δ/γ selectivity, as a comparison of 45 and quinazolinone 109 indicates. They also published a detailed treatise on the relative merits of the PI3K δ inhibitor 47, 109, and pan-inhibitor 10 (PIK-90). By use of 2 different human cell lines, 7 stimulating agents, and up to 20 different expressed proteins known to be involved in inflammation, they found that the greatest suppression of these cytokines and other inflammatory second messengers came from the dual PI3K δ/γ inhibitor 109. They also compared these results to several known antiinflammatory agents and found that 109 most closely matches prednisolone in a functional way. On the basis of this in vitro treatment, the team concluded that the targeting of RA and other diseases of inflammation would be most effective with a dual PI3K δ/γ inhibitor such as **109**.¹⁸¹ Takeda/Intellikine is championing the development of a dual PI3K γ and PI3K δ inhibitor for inflammatory diseases and hematologic malignancies.⁷⁵ They disclose at least nine compounds, exemplified by thienopyrimidinone 110,¹⁸² with PI3K δ and PI3K γ potency

of less than 100 nM in enzyme assays. The compounds were weakly potent (>10 μ M) toward PI3K α and PI3K β but active with EC₅₀ < 100 nM in a B-cell proliferation assay. The structures all contain a phenolic substituted pyrazolopyrimidine side chain as found in **45** and **46**.

Takeda/Intellikine has put forward two dual PI3K γ/δ inhibitors. INK055 (structure not disclosed) was dosed therapeutically at 30 mg/kg in a CIA model of RA with clinical scoring >2-fold lower than vehicle control at day 34 (dosing started on day 30). Interestingly, anti type II collagen antibody levels were unaffected, consistent with a PI3K γ driven response.^{67,109} Prior to their acquisition by Takeda, Intellikine licensed the dual PI3K γ/δ inhibitor IPI-145 (formerly INK1197, structure not disclosed) to Infinity Pharmaceuticals Inc., which is currently in phase I for hematological malignancies.¹⁸³

STRUCTURAL ASPECTS OF THE PI3Kδ AND PI3Kγ INHIBITORS

Development of class I PI3K isoform specific inhibitors is challenging because of the high sequence identity exhibited between catalytic p110 subunits and the even higher level of conservation of amino acids within the ATP-binding site (Table 8).¹⁸⁴ However, as the preceding sections demonstrate,

Table 8. Amino Acid Sequence Identity (%) between Human PI3K Isoforms

kinase domain	p110δ	p110γ	p110α	p110β
p110δ		42	47	73
p110γ			44	43
p110α				48
active site	p110δ	p110γ	p110α	p110β
p110δ		68	62	81

inhibitors with varying isoform selectivities exist, and indeed, some are quite selective. Central to our understanding of the structure of the p110 proteins and of their complexes with small molecule ATP-competitive inhibitors were the publications of the crystal structures of porcine and human class IB p110 γ proteins.^{185,186}

These structures elucidated the key features of the active site and provided detailed views of how classic pan-PI3K inhibitors such as 1 and 2 inhibited the PI3Ks. Briefly, inhibitors bound to the active site by making one or more hydrogen bonds to residues in the hinge region of the kinase domain, analogous to interactions made by ATP-competitive inhibitors of protein kinases. Hydrogen bonds in the hinge region are often the strongest contact that ATP competitive inhibitors make, estimated to account for 40-60% of the overall binding energy for inhibitors of certain protein kinases.¹⁸⁷ Most PI3K inhibitors also interact with the large, flat hydrophobic face of a conserved tyrosine residue (Tyr867 in p110 γ), and many extend back into the affinity pocket where additional hydrophobic interactions can be gained to boost potency and where, with the presence of an appropriate heteroatom, inhibitors can form hydrogen bonds with the catalytic lysine (Lys833 in p110 γ) or other hydrophilic residues.³⁴ In the case of 2 (Figure 10A), the morpholine ring hydrogen-bonds to hinge residue Val882 and the edge of the morpholine ring interacts with the face of Tyr867. The chromanone carbonyl group points toward the affinity pocket, and the phenyl group sitting orthogonal to the chromanone ring occupies the ribose

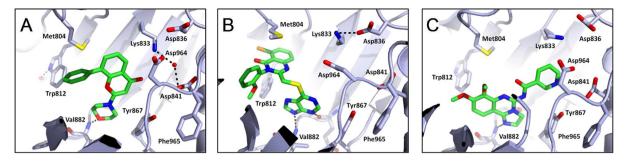


Figure 10. (A) Crystal structure of porcine p110 γ and 2 (PDB code 1E7V). (B) Crystal structure of human p110 γ and 4 (PDB code 2CHW), a propeller-shaped inhibitor. (C) Crystal structure of human p110 γ and 10 (PDB code 2CHX), a flat inhibitor.

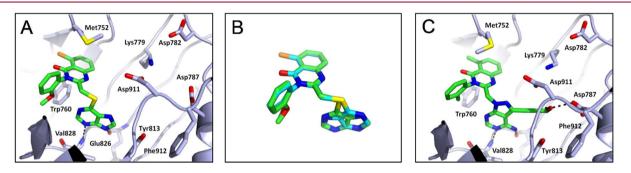


Figure 11. (A) Crystal structure of murine p110 δ and 4 (PDB code 2WXF). (B) Overlay of 4 from murine p110 δ (green) and human p110 γ (blue). The purine binding mode depicted in green is likely the energetically more favorable conformation for both isoforms. (C) Crystal structure of murine p110 δ and 47 (PDB code 2WXI) showing an alkynol group in the affinity pocket.

pocket. Subsequent work by numerous groups has resulted in the structure elucidation of many additional p110 γ crystal structures in complex with small molecule inhibitors. More recently, crystal structures of the class IA p110 α , p110 β , and p110 δ enzymes have been published.^{111,188–191} The availability of multiple inhibitor-bound p110 δ crystal structures in particular has allowed for the detailed analysis of binding interactions of pan- and selective class I P13K inhibitors.

Crystal structure analysis combined with data from biochemical and cellular assays can be used to understand the molecular basis for the observed inhibitor selectivities. In notable work,³⁴ extensive biochemical profiling of various inhibitor chemotypes combined with crystal structures of three distinct molecules in PI3Ky allowed the authors to correlate and explain the observed selectivities for various structural classes of PI3K inhibitors. The diverse inhibitors clustered into several different classes but bound to $p110\gamma$ in one of two principal binding modes: (I) a propeller shaped binding mode exemplified by quinazolinones 3 and 4 (Figure 10B) and (II) a flatter, more classical kinase binding mode exemplified by 10 (Figure 10C). There is a distinct difference in the overall selectivity between these binding modes. Flat inhibitors such as 10 can occupy the binding pocket without any significant amino acid rearrangements. They extend into the affinity pocket (10 uses a pyridine ring), and they are typically pan-PI3K inhibitors. On the other hand, PI3K δ selective quinazolinone 3 binds only when a key methionine residue (Met804 PI3K γ , conserved in all isoforms) adopts an alternative conformation that opens a hydrophobic pocket (specificity pocket) sitting orthogonal to the position of residues in the p110 γ hinge region (Figure 10B). This ligandinduced methionine reorientation, first observed in the crystal structure of $p110\gamma$ and 4, also induces a shift of the peptide backbone that propagates through the adjacent loop (equivalent to the P-loop in protein kinases, amino acids 803–811 in PI3K γ). It was postulated that this conformational change, although possible in all class I PI3Ks, was most facile in PI3K δ , thereby accounting for the higher binding affinity of propeller shaped inhibitors on p110 δ relative to the other isoforms. Following publication of the first p110 α crystal structure, an examination of 4 binding to this isoform concluded that differences in the conformation of the P-loop precluded binding of 4 because of steric reasons, even with a conformational change in Met772.¹⁹² However, a recent crystal structure of murine p110 α showing a ligand-induced Met772 reorientation calls that conclusion into question.¹⁸⁹ In addition to the crystallographic work, computational approaches have also been used to examine issues of PI3K inhibitor selectivity, and they have reached varying conclusions.

Crystal structures of both flat and propeller-shaped molecules in p110 δ have reinforced and extended the initial selectivity arguments. Crystal structures of the most selective analogues, 3 and 4 (Figure 11), provide an explanation of selectivity that is similar but perhaps more thorough than what was argued based on the PI3K γ structures. The N-terminal lobe of the kinase domain in p110 proteins contains a loop spanning the k α 1 and k α 2 helices. This loop is longer in PI3K γ than in PI3K δ , and it sits on top of the P-loop, limiting its flexibility and leading to less facile rearrangement of the active site in PI3K γ relative to PI3K δ . Molecular dynamics simulations based on the crystal structures demonstrated a lower degree of flexibility in p110 γ versus p110 δ , thus supporting this argument. Also contributing to lowered flexibility in p110 γ is a hydrogen bond network that restricts the movement of Trp812, an essential component of the hydrophobic pocket into which the propeller shaped molecules bind. The nearly identical conformation of 4 in p110 δ versus p110 γ (Figure 11B) further bolsters the argument that differences in the

ability of the active sites to accommodate the inhibitor account for potency differences rather than an altered binding conformation of the inhibitor in each isoform. The PI3K δ crystal structures also demonstrated that simultaneously occupying the selectivity and affinity pockets is possible, and indeed, propeller shaped analogues 42, 43, 44, 46, 47, and 109 take advantage of this strategy to impart an additional level of potency toward PI3K δ . For example, in 47 the pendent alkynol hydrogen-bonds to Asp787 and to the backbone NH of Asp911(Figure 11C), resulting in significantly improved potency relative to 3 and 4. Even among the more selective propeller shaped molecules, however, a divergence of selectivity is obtainable by only minor modification. For example, 46 forms hydrogen bonds between the phenol -OH group and Tyr813 and between the meta-fluoro substituent and Lys779, whereas 109, a constitutional isomer of 46, forms only the hydrogen bond with Lys779. As a consequence, 46 is a more PI3K δ selective inhibitor than **109**. This example highlights the fact that although propeller shaped compounds can be very PI3K δ selective, care must be taken to monitor the selectivity of such compounds, as variations are made on the periphery of the molecule.

Among the many crystal structures of p110 γ now available in the public realm, only a few contain inhibitors reported to be PI3K γ selective. The structures of **6** and **8** from Serono provide insight into the structural features that promote selectivity for PI3K γ over the other isoforms. Although PI3K γ can be inhibited by propeller shaped molecules as discussed above, such molecules seemed destined to carry along PI3K δ activity. In contrast, the Serono structures show compact molecules that form a hydrogen bond with hinge residue Val882 and extend toward the affinity pocket where they form an additional hydrogen bond to catalytic Lys833 using their thiazolidinedione moieties (Figure 12). These modestly sized molecules appear

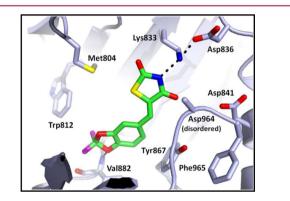


Figure 12. Crystal structure $p110\gamma$ -selective inhibitor 8 and human $p110\gamma$ (PDB code 2A4Z).

to have the optimal dimensions to span this distance when Lys833 is in a conformation that sits lower in the affinity pocket, constricting its size. A computational analysis of the selectivity of these molecules using homology models of class IA PI3Ks reinforced the idea that these compounds are ideally sized for PI3K γ , as identical binding interactions were not reproduced in the other isoforms.¹⁹⁴ A separate series of PI3K γ selective inhibitors reported recently follows this paradigm.¹⁷⁰ From the crystal structures of the class IA enzymes, it appears that the kinase domain conformation may also play a role in modulating the affinity of these molecules against various isoforms, as p110 δ is distinct from p110 α and p110 γ in the

orientation of its N-lobe with respect to its C-lobe in the kinase domain and molecules **6**, **8**, and **9** are significantly less potent on PI3K δ . By the same argument, it seems likely that achieving selectivity over PI3K α will be the most challenging aspect of designing PI3K γ selective inhibitors, an idea supported by the inhibitor profiling already reported³⁴ and borne out in a recently published work.¹⁷⁰

Whereas the selectivity profiles of many PI3K inhibitors are easy to understand based on our current knowledge of class I PI3K inhibitor binding modes, rational design of exquisitely selective PI3K inhibitors still presents a challenge. Future work is sure to produce further insights into mechanisms of achieving isoform selective inhibitors, as the recent publication of hexahydroquinoxalinone compound 58 illustrates.¹¹¹ This compound is PI3K δ selective and yet does not adopt a propeller shape. The crystal structure of **58** in p110 δ shows that rather than sitting sandwiched between the Met752 and Trp760 in the selectivity pocket, the compound sits adjacent to these two amino acids in a small pocket formed by Thr750. The other isoforms do not have such a pocket because the residue at this position is a much larger arginine or lysine. Indeed, the recent advances in molecular biology and protein expression that enabled the crystallization of the class IA isoforms may provide further novel insights into the selectivities of PI3K inhibitors as more structures are solved.^{188,189,195}

DISCUSSION AND CONCLUSION

Teasing out the differences that PI3K δ and PI3K γ have in the immune system is not a trivial endeavor. Clearly the adaptive immune system has evolved in proximity to the innate system, and this relationship is illustrated by the interplay between these two key kinases. For example, both PI3K δ and PI3K γ over multiple steps influence neutrophil responses to sites of infection or inflammation such that in diseases that activate this cell type, targeting one or the other of these two kinases could prove advantageous. Mast cell degranulation is affected by both PI3K γ and PI3K δ , as has been shown in several genetic and pharmacologic studies. Conflicting data (IgE levels, tracheal smooth muscle hyperresponsiveness) in some asthma model studies are indicative of overlapping outcomes, in spite of the distinct functions of PI3K δ and PI3K γ . The inhibition of PI3K δ has a great impact on B-cell proliferation, antibody production, and Ig class switching, but the inhibition of PI3K γ has negligible effects on B-cells. This is an advantage for drugs targeting PI3K δ for diseases that have a significant B-cell component such as shown to be the case for RA. Leukocyte infiltration is a hallmark of asthma, ALI/ARDS, IPF, RA, and colitis (IBD), and PI3K γ controls much of this chemotaxis. A case can be made for targeting RA with PI3K γ inhibitors by inhibition of leukocyte migration. Also, a PI3K γ inhibitor was effective in a lupus model over several months and in a MS model, but no such data have been described for PI3K δ inhibitors. Dual PI3K γ and PI3K δ inhibitors have been getting increasing scrutiny, although the data from animal models rely mainly on 7. The use of this compound in asthma and COPD models showed promise. In spite of the deleterious outcomes of the dual PI3K γ KO and PI3K δ KDKI mice, the severe lymphopenia and other adverse effects were not replicated in the study involving PI3K γ KO mice and 3. Instead, there was a further reduction in the mean ankle thickness and an improved clinical score relative to 3 dosed WT mice.

It is tempting to make judgments of the utility of the two targets based on the number of studies reported for the various

animal disease models. Using this approach, researchers have been interested in selective PI3K δ inhibitors for asthma, COPD, and selective PI3Ky inhibitors for IBD, SLE, and RA. The sheer number of reported studies targeting PI3Ky would indicate that this is the preferred target, but this is refuted somewhat by the dearth of chemical matter related to this target. There are many more patent applications for PI3K δ inhibitors than for PI3K γ inhibitors, although many PI3K γ inhibitors could be found in patents that are more closely aligned to mTOR as the dominant target. In this review we have ignored for the most part the vast IP space that mentions mTOR as co-target or PI3K α for that matter. Finally, the prevalence of PI3K δ inhibitors verses PI3K γ inhibitors could simply be due to the relative ease of finding isoform and kinase selective PI3K δ inhibitors, as the structural data indicate. Dual inhibitors are less frequently mentioned in the literature and are possibly masked in related patents, as most applications are written to cover all isoforms. Interestingly, there are several dual inhibitors that have advanced toward the clinic.

A perusal of the selectivity data in Tables 4 and 5 indicates that the propeller shape seems to dominate inhibitor design. Of the 18 compounds with PI3K γ /PI3K δ selectivity of greater than 20, 11 are of this type. The remaining seven compounds, while all having PI3K γ /PI3K δ selectivity of greater than 20, fall considerably short in the selectivity over the other isoforms (see, for example, 12, 55, and 57). On the other hand, 9 of the 11 propeller shaped analogues exhibit PI3K δ selectivity over the other isoforms of greater than 100-fold. However, as was pointed out earlier, the Serono compound 58 is not propeller shaped and does have similar selectivity compared to these compounds. Whether these structural data have been utilized in further inhibitor design is unknown. Roche/Genentech has shown that the more flat molecules such as 14-17, while structurally very similar to 12, have considerably higher selectivity over PI3K α and may have selectivity over PI3K γ . Less can be said about the PI3K γ inhibitor class, as there are fewer published reports dealing with PI3Ky-selective compounds, although the best examples of these are relatively compact molecules that do not include any propeller shaped compounds.

One paradox of drug discovery is that the initial biological hypothesis is of necessity based on limited understanding of complex systems. Fundamental biological research is facilitated by progress in the development of small molecule tools that help address understanding of target coverage and selectivity requirements as well as issues such as target redundancy or compensation. As this understanding develops, the drug product profile may change considerably from that initially proposed on the basis of a one target-one effect model. Additionally, animal disease models mentioned in this review have uncertain relevance to actual human disease states and their utility as predictors of clinical success is in question. The question of final target validation is likely to remain unanswered until one or more compounds targeting either PI3K γ or PI3K δ complete clinic trials for diseases of autoimmunity/inflammation.

AUTHOR INFORMATION

Corresponding Author

*Phone: 1-650-224-2406. E-mail: tcushing@amgen.com.

Notes

The authors declare the following competing financial interest(s): The authors are all employees of Amgen Inc.

Biographies

Timothy D. Cushing obtained his B.Sc. degree from the University of Minnesota and received his Ph.D. degree in Synthetic Organic Chemistry from Colorado State University in 1993 where he completed the total synthesis of the natural product paraherquamide B under the direction of Professor Robert M. Williams. After an NIH postdoctoral fellowship with Professor Gregory L. Verdine at Harvard University, MA, he joined Tularik in 1995 where he worked on a variety of projects focused on viral, inflammation, and oncology targets. In 2004 he began working for Amgen and continued in the area of inflammation and oncology, including leading the chemistry efforts related to PI3K δ . He is currently a Principal Scientist in the Department of Therapeutic Discovery at Amgen in South San Francisco, CA.

Daniela P. Metz received her B.Sc. in Parasitology from the University College of North Wales, UK, in 1987. This was followed by an M.Sc. (1989) and Ph.D. (1993) in Immunology from the University of Birmingham, U.K., and University College of London, U.K., respectively. From 1994 to 1999 she was a Postdoctoral Fellow in the lab of Professor Kim Bottomly at Yale University Medical School's Section of Immunobiology. Dr. Metz remained at Yale as an Associate Research Scientist (1998–2000) before she moved to the University of Rochester, NY, as a Research Assistant Professor in the Department Microbiology and Immunology. In 2003 she joined Amgen, where she is currently, as a Director of Research in the Inflammation Research Group, leading various immunological projects, including the PI3K δ program.

Douglas A. Whittington received his B.Sc. in Chemistry from the University of Michigan in 1994 and his Ph.D. in Inorganic Chemistry from the Massachusetts Institute of Technology in 2000, where he worked under the direction of Professor Stephen J. Lippard. As a Postdoctoral Fellow in Professor David W. Christianson's lab at the University of Pennsylvania, he solved crystal structures of enzymes involved in monoterpenoid biosynthesis and zinc-mediated hydrolysis, including the antibiotic target LpxC. Since joining Amgen in 2004, he has worked on structure-based drug design projects in the areas of oncology, inflammation, and neuroscience. He is currently a Principal Scientist in the Department of Molecular Structure and Characterization at Amgen in Cambridge, MA.

Lawrence R. McGee received his B.Sc. in Chemistry from the University of Utah in 1974 and his Ph.D. in Organic Chemistry from Caltech, CA, in 1982, where he worked with Professor David A. Evans. He began his industrial career at DuPont Central Research followed by experience in drug discovery at Genentech, Gilead Sciences, and Tularik. Since joining Amgen in 2004, he has worked on drug discovery projects in the areas of diabetes, oncology, inflammation, and neuroscience. He is currently a Scientific Director in the Department of Therapeutic Discovery at Amgen in South San Francisco, CA.

ACKNOWLEDGMENTS

Thanks are given to Julio Medina for useful discussions and encouragement and to Jennifer Seganish for the preparation of Figure 1.

ABBREVIATIONS USED

ADME, absorption, distribution, metabolism, and excretion; AHR, airway hyperresponsiveness; AIA, antigen-induced arthritis; ALI/ARDS, acute lung injury/acute respiratory distress syndrome; AML, acute myleloid leukemia; APC, antigen presenting cells; ATP, adenosine triphosphate; B1, a class of B cell lymphocytes; BALF, bronchoalveolar lavage fluid; BCR, B-cell receptor; b.i.d., twice a day; BLM, bleomycin; BMMC, bone-marrow-derived mast cell; BTK, Bruton's tyrosine kinase; cAMP, cyclic adenosine monophosphate; CCL2, chemokine (C-C motif) ligand 2; CCL5, chemokine (C-C motif) ligand 5; CD, cluster of differentiation; CIA, collagen induced arthritis; Cl, clearance; CLL, chronic lymphocytic leukemia; CNS, central nervous system; COPD, chronic obstructive pulmonary disease; DNA-PK, DNAdependent protein kinase; DNP, dinitrophenyl; DSS, dextran sodium sulfate; ETK, cryptic autophosphorylating protein tyrosine kinase; fMLP, N-formylmethionylleucylphenylalanine; FO, follicular; FOXO3a, forkhead box O3; GPCR, G-proteincoupled receptor; GR- α , glutocorticoid receptor α ; HWB, human whole blood; IBD, inflammatory bowel disease; IFN- γ , interferon γ ; Ig, immunoglobulin; IL, interleukin; IPF, Idiopathic pulmonary fibrosis; ITK, interleukin-2-inducible Tcell kinase; KDKI, kinase dead knock-in; KO, knockout; LPS, lipopolysaccharide; MI, myocardial infarction; MMP3, matrix metalloproteinase 3; MOG, myelin oligodendrocyte glycoprotein; MS, multiple sclerosis; MTD, maximum tolerated dose; mTOR, mammalian target of rapamycin; MZ, marginal zone; NHL, non-Hodgkin's lymphoma; OVA, ovalbumin; PCA, passive cutaneous anaphylaxis; PBMC, peripheral blood mononuclear cell; PDGF, platelet-derived growth factor; PDK1, phosphoinositide dependent kinase 1; PIKK, phosphatidylinositol 3-kinase-related kinase; PKB/Akt, protein kinase B; PMN, polymorphonuclear leukocyte (neutrophil); PTEN, phosphatase and tensin homologue; q.d., once a day dosing; RA, rheumatoid arthritis; RTK, receptor tyrosine kinase; ROS, reactive oxygen species; SCW, Sstreptococcal cell wall; SF, synovial fibroblast; SLE, systemic lupus erythematosus; TCR, T-cell receptor; T_h1, T helper cell type 1; T_h2, T helper cell type 2; TLR, Toll-like receptor; TNF- α , tumor necrosis factor α ; T_{reg}, T regulatory cell; VEGF, vascular endothelial growth factor; VV, vaccinia virus; WT, wild type

REFERENCES

(1) Kong, D.; Yamori, T. Advances in Development of Phosphatidylinositol 3-Kinase Inhibitors. *Curr. Med. Chem.* 2009, 16, 2839–2854.

(2) Ameriks, M.; Venable, J. D. Small Molecule Inhibitors of Phosphoinositide 3-Kinase (PI3K) δ and γ . *Curr. Top. Med. Chem.* **2009**, 738–753.

(3) Sundstrom, T. J.; Anderson, A. C.; Wright, D. L. Inhibitors of Phosphoinositide-3-kinase: A Structure-Based Approach to Understanding Potency and Selectivity. *Org. Biomol. Chem.* **2009**, *7*, 840–850.

(4) Parker, P. J. The Ubiquitous Phosphoinositides. *Biochem. Soc. Trans.* 2004, 32, 893–898.

(5) Hawkins, P. T.; Anderson, K. E.; Davidson, K.; Stephens, L. R. Signaling through Class 1 PI3Ks in Mammalian Cells. *Biochem. Soc. Trans.* **2006**, *34*, 647–662.

(6) Fruman, D. A.; Bismuth, G. Fine Tuning the Immune Response with PI3K. *Immunol. Rev.* **2009**, *228*, 253–272.

(7) McInnes, I. B.; Schett, G. Cytokines in the Pathogenesis of Rheumatoid Arthritis. *Nat. Rev. Immunol.* **2007**, *7*, 429–442.

(8) Mount, G. R.; Gilliland, W. R. Emerging Biological Therapies in Systemic Lupus Erythematosus. *Clin. Pharmacol. Ther.* **2008**, *83*, 167–171.

(9) Compston, A.; Coles, A. Multiple Sclerosis. Lancet 2008, 372, 1502–1517.

(10) Lowes, M. A.; Bowcock, A. M.; Krueger, J. G. Pathogenesis and Therapy of Psoriasis. *Nature* **2007**, *445*, 866–873. (11) Decramer, M.; Janssens, W.; Miravitlles, M. Chronic Obstructive Pulmonary Disease. *Lancet* **2012**, *379*, 1341–1351.

(12) Reutershan, J.; Saprito, M. S.; Wu, D.; Rückle, T.; Ley, K. Phosphoinositide 3- Kinase γ Required for Lipopolysaccharide-Induced Transepithelial Neutrophil Trafficking in the Lung. *Eur. Respir. J.* **2010**, *35*, 1137–1147.

(13) Holgate, S. T.; Davies, D. E. Rethinking the Pathogenesis of Asthma. *Immunity* **2009**, *31*, 362–367.

(14) King, T. E., Jr.; Pardo, A.; Selman, M. Idiopathic Pulmonary Fibrosis. Lancet 2011, 378, 1949–1961.

(15) Xavier, R. J.; Podolsky, D. K. Unravelling the Pathogenesis of Inflammatory Bowel Disease. *Nature* **2007**, *448*, 427–434.

(16) Hori, M.; Nishida, K. Oxidative Stress and Left Ventricular Remodelling after Myocardial Infarction. *Cardiovasc. Res.* **2009**, *81*, 457–464.

(17) Bonjouklian, R.; Vlahos, C. J.; Powis, G. Inhibition of Phosphatidylinositol 3-Kinase with Wortmannin and Its Analogs. U.S. Patent 5,378,725, Jan 3, 1995.

(18) Vlahos, C. J.; Matter, W. F.; Hui, K. Y.; Brown, R. F. A Specific Inhibitor of Phosphatidylinositol 3-Kinase, 2-(4-Morpholinyl)-8-phenyl-4H-1-benzopyran-4-one (LY294002). *J. Biol. Chem.* **1994**, 269, 5241–5248.

(19) Sadhu, C.; Dick, K.; Treiberg, J.; Sowell, C. G.; Kesicki, E. A.; Oliver, A. Quinazolinone Derivatives as Inhibitors of Human Phosphatidylinositol 3-Kinase delta. WO2001081346A2, 2001.

(20) Pomel, V.; Klicic, J.; Covini, D.; Church, D. D.; Shaw, J. P.; Roulin, K.; Burgat-Charvillon, F.; Valognes, D.; Camps, M.; Chabert, C.; Gillieron, C.; Francon, B.; Perrin, D.; Leroy, D.; Gretener, D.; Nichols, A.; Vitte, P. A.; Carboni, S.; Rommel, C.; Schwarz, M. K.; Rückle, T. Furan-2-ylmethylene thiazolidinediones as Novel, Potent, and Selective Inhibitors of Phosphoinositide 3-Kinase γ. J. Med. Chem. **2006**, 49, 3857–3871.

(21) Camps, M.; Rückle, T.; Ji, H.; Ardissone, V.; Rintelen, F.; Shaw, J.; Ferrandi, C.; Chabert, C.; Gillieron, C.; Francon, B.; Martin, T.; Gretener, D.; Perrin, D.; Leroy, D.; Vitte, P. A.; Hirsch, E.; Wymann, M. P.; Cirillo, R.; Schwarz, M. K.; Rommel, C. Blockade of PI3K γ Suppresses Joint Inflammation and Damage in Mouse Models of Rheumatoid Arthritis. *Nat. Med.* **2005**, *11*, 936–943.

(22) Wrasidlo, W.; Doukas, J.; Royston, I.; Noronha, G.; Hood, J. D.; Dneprovskaia, E.; Gong, X.; Splittgerber, U.; Zhao, N. Preparation of Vasculostatic Agents and Methods of Use. WO2004030635A2, 2004.

(23) Okkenhaug, K.; Bilancio, A.; Farjot, G.; Priddle, H.; Sancho, S.; Peskett, E.; Pearce, W.; Meek, S. E.; Salpekar, A.; Waterfield, M. D.; Smith, A. J. H.; Vanhaesebroeck, B. Impaired B and T Cell Antigen Receptor Signaling in p110 delta PI 3-Kinase Mutant Mice. *Science* **2002**, *297*, 1031–1034.

(24) Hirsch, E.; Katanaev, V. L.; Garlanda, C.; Azzolino, O.; Pirola, L.; Silengo, L.; Sozzani, S.; Mantovani, A.; Altruda, F.; Wymann, M. P. Central Role for G Protein-Coupled Phosphoinositide 3-Kinase γ in Inflammation. *Science* **2000**, *287*, 1049–1053.

(25) Patrucco, E.; Notte, A.; Barberis, L.; Selvetella, G.; Maffei, A.; Brancaccio, M.; Marengo, S.; Russo;, G.; Azzolino, O.; Rybalkin, S. D.; Silengo, L.; Altruda, F.; Wetzker, R.; Wymann, M. P.; Lembo, G.; Hirsch, E. PI3K γ Modulates the Cardiac Response to Chronic Pressure Overload by Distinct Kinase-Dependent and -Independent Effects. *Cell* **2004**, *118*, 375–387.

(26) Ji, H.; Rintelen, F.; Waltzinger, C.; Meier, D. B.; Bilancio, A.; Pearce, W.; Hirsch, E.; Wymann, M. P.; Rückle, T.; Camps, M.; Vanhaesebroeck, B.; Okkenhaug, K.; Rommel, C. Inactivation of PI3K γ and PI3K δ Distorts T-Cell Development and Causes Multiple Organ Inflammation. *Blood* **2007**, *110*, 2940–2947.

(27) Note on the nomenclature for this article: PI3K γ or PI3K δ knockout (KO) refers to a genetically modified animal (mouse) or its cells where the entire p110 γ or p110 δ catalytic subunit of the corresponding PI3K heterodimer has been deleted. This is in contrast to the PI3K γ or PI3K δ kinase dead knock-in (KDKI) where a point mutation has been genetically engineered into the ATP binding pocket of the corresponding p110 catalytic subunit abrogating the enzyme activity. The scaffolding function of this type is presumably intact.

(28) Wright, H. L.; Moots, R. J.; Bucknall, R. C.; Edwards, S. W. Neutrophil Function in Inflammation and Inflammatory Diseases. *Rheumatology* **2010**, *49*, 1618–1631.

(29) Edwards, J. C. W.; Cambridge, G. Sustained Improvement in Rheumatoid Arthritis following a Protocol Designed To Deplete B Lymphocytes. *Rheumatology* **2001**, *40*, 205–211.

(30) Yanaba, K.; Hamaguchi, Y.; Venturi, G. M.; Steeber, D. A., St.; Clair, E. W.; Tedder, T. F. B Cell Depletion Delays Collagen-Induced Arthritis in Mice: Arthritis Induction Requires Synergy between Humoral and Cell-Mediated Immunity. *J. Immunol.* **200**7, *179*, 1369– 1380.

(31) (a) Li, J; Kuzin, I; Moshkani, S; Proulx, S. T.; Xing, L; Skrombolas, D.; Dunn, R.; Sanz, I; Schwarz, E. M.; Bottaro, A. Expanded CD23+/CD21hi B Cells in Inflamed Lymph Nodes Are Associated with the Onset of Inflammatory–Erosive Arthritis in TNF-Transgenic Mice and Are Targets of Anti-CD20 Therapy. *J. Immunol.* **2010**, *184*, 6142–6150. (b) Rituximab was found to be ineffective in a large scale phase II/III clinical trial for SLE.

(32) Vallerskog, T.; Gunnarsson, I.; Widhe, M.; Risselada, A.; Klareskog, L.; van Vollenhoven, R.; Malmström, V.; Trollmo, C. Treatment with Rituximab Affects Both the Cellular and the Humoral Arm of the Immune System in Patients with SLE. *Clin. Immunol.* 2007, 122, 62–74.

(33) Ferrandi, C.; Ardissone, V.; Ferro, P.; Rückle, T.; Zaratin, P.; Ammannati, E.; Hauben, E.; Rommel, C.; Cirillo, R. Phosphoinositide 3-Kinase γ Inhibition Plays a Crucial Role in Blocking the Early Steps of Inflammation by Blocking Neutrophil Recruitment. *J. Pharmacol. Exp. Ther.* **2007**, *322*, 923–930.

(34) Knight, Z. A.; Gonzalez, B.; Feldman, M. E.; Zunder, E. R.; Goldenberg, D. D.; Williams, O.; Loewith, R.; Stokoe, D.; Balla, A.; Toth, B.; Balla, T.; Weiss, W. A.; Williams, R. L.; Shokat, K. M. A Pharmacological Map of the PI3-K Family Defines a Role for P110 α in Insulin Signaling. *Cell* **2006**, *125*, 733–747.

(35) Palanki, M. S. S.; Dneprovskaia, E.; Doukas, J.; Fine, R.,M.; Hood, J.; Kang, X.; Lohse, D.; Martin, M.; Noronha, G.; Soll, R. M.; Wrasidlo, W.; Yee, S.; Zhu, H. Discovery of 3,3'-(2,4-Diaminopteridine-6,7-diyl)diphenol as an Isozyme-Selective Inhibitor of PI3K for the Treatment of Ischemia Reperfusion Injury Associated with Myocardial Infarction. J. Med. Chem. 2007, 18, 4279–4294.

(36) Soond, D. R.; Bjørgo, E.; Moltu, K.; Dale, V. Q.; Patton, D. T.; Torgerson, K. M.; Galleway, F.; Twomey, B.; Clark, J.; Gaston, J. S. H.; Taskén, K.; Bunyard, P.; Okkenhaug, K. PI3K P110 δ Regulates T-Cell Cytokine Production During Primary and Secondary Immune Responses in Mice and Humans. *Blood* **2010**, *115*, 2203–2213.

(37) Sasaki, T.; Irie-Sasaki, J.; Jones, R. G.; Oliveira-dos-Santos, A. J.; Stanford, W. L.; Bolon, B.; Wakeham, A.; Itie, A.; Bouchard, D.; Kozieradzki, I.; Joza, N.; Mak, T. W.; Ohashi, P. S.; Suzuki, A.; Penninger, J. M. Function of PI3K γ in Thymocyte Development, T Cell Activation, and Neutrophil Migration. *Science* **2000**, 287, 1040– 1046.

(38) Alcázar, I.; Marqués, M.; Kumar, A.; Hirsch, E.; Wymann, M.; Carrera, A. C.; Barber, D. F. Phosphoinositide 3-Kinase γ Participates in T cell Receptor-Induced T Cell Activation. *J. Exp. Med.* **2007**, *204*, 2977–2987.

(39) Curnock, A. P.; Sotsios, Y.; Wright, K. L.; Ward, S. G. Optimal Chemotactic Responses of Leukemic T Cells to Stromal Cell-Derived Factor-1 Requires the Activation of Both Class 1A and Class 1B Phosphoinositide 3-Kinases. J. Immunol. **2003**, *170*, 4021–4030.

(40) Smith, L. D.; Hickman, E. S.; Parry, R. V.; Westwick, J.; Ward, S. G. PI3K γ Is the Dominant Isoform Involved in Migratory Responses of Human T Lymphocytes: Effects of ex Vivo Maintenance and Limitations of Non-Viral Delivery of siRNA. *Cell. Signalling* **2007**, *19*, 2528–2539.

(41) Sadhu, C.; Masinovsky, B.; Dick, K.; Sowell, C. G.; Staunton, D. E. Essential Role of Phosphoinositide 3-Kinase δ in Neutrophil Directional Movement. *J. Immunol.* **2003**, *170*, 2647–2654.

(42) Puri, K. D.; Doggett, T. A.; Douangpanya, J.; Hou, Y.; Tino, W. T.; Wilson, T.; Graf, T.; Clayton, E.; Turner, M.; Hayflick, J. S.; Diacovo, T. G. Mechanism and Implications of Phosphoinositide 3-

Kinase delta in Promoting Neutrophil Trafficking into Inflamed Tissue. *Blood* **2004**, *103*, 3448–3456.

(43) Ali, K.; Bilancio, A.; Thomas, M.; Pearce, W.; Gilfillan, A. M.; Tkaczyk, C.; Kuehn, N.; Gray, A.; Giddings, J.; Peskett, E.; Fox, R.; Bruce, I.; Walker, C.; Sawyer, C.; Okkenhaug, K.; Finan, P.; Vanhaesebroeck, B. Essential Role for the P110 δ Phosphoinositide 3-Kinase in the Allergic Response. *Nature* **2004**, 431, 1007–1011.

(44) Ali, K.; Camps, M.; Pearce, W. P.; Ji, H.; Rückle, T.; Kuehn, N.; Pasquali, C.; Chabert, C.; Rommel, C.; Vanhaesebroeck, B. Isoform-Specific Functions of Phosphoinositide 3-Kinases: PI3K δ but not PI3K γ Promotes Optimal Allergic Responses in Vivo. *J. Immunol.* **2008**, 180, 2538–2544.

(45) Laffargue, M.; Calvez, R.; Finan, P.; Trifilieff, A.; Barbier, M.; Altruda, F.; Hirsch, E.; Wymann, M. P. Phosphoinositide 3-Kinase γ Is an Essential Amplifier of Mast Cell Function. *Immunity* **2002**, *16*, 441–451.

(46) Thomas, M.; Edwards, M. J.; Sawicka, E.; Duggan, N.; Hirsch, E.; Wymann, M. P.; Owen, C.; Trifilieff, A.; Walker, C.; Westwick, J.; Finan, P. Essential Role of Phosphoinositide 3-Kinase gamma in Eosinophil Chemotaxis within Acute Pulmonary Inflammation. *Immunology* **2009**, *126*, 413–422.

(47) Takeda, M.; Ito, W.; Tanabe, M.; Ueki, S.; Kato, H.; Kihara, J.; Tanigai, T.; Chiba, T.; Yamaguchi, K.; Kayaba, H.; Imai, Y.; Okuyama, K.; Ohno, I.; Sasaki, T.; Chihara, J. Allergic Airway Hyperresponsiveness, Inflammation, and Remodeling Do Not Develop in Phosphoinositide 3-Kinase γ -Deficient Mice. *J. Allergy Clin. Immunol.* **2009**, *123*, 805–812.

(48) Kang, B. N.; Ha, S. G.; Ge, X. N.; Hosseinkhani, M. R.; Bahaie, N. S.; Greenberg, Y.; Blumenthal, M. N.; Puri, K. D.; Rao, S. P.; Sriramarao, P. The p110 δ Subunit of PI3K Regulates Bone Marrow-Derived Eosinophil Trafficking and Airway Eosinophilia in Allergen-Challenged Mice. *Am J. Physiol.: Lung Cell. Mol. Physiol.* 2012, 302, 1179–1191.

(49) Doukas, J.; Eide, L.; Stebbins, K.; Racanelli-Layton, A.; Dellamary, L.; Martin, M.; Dneprovskaia, E.; Noronha, G.; Soll, R.; Wrasidlo, W.; Acevedo, L. M.; Cheresh, D. A. Aerosolized Phosphoinositide 3-Kinase γ/δ Inhibitor TG100-115 [3-[2,4-Diamino-6-(3-hydroxyphenyl)-pteridin-7-yl]phenol] as a Therapeutic Candidate for Asthma and Chronic Obstructive Pulmonary Disease. *J. Pharmacol. Exp. Ther.* **2009**, *328*, 758–765.

(50) Ghigo, A.; Damilano, F.; Braccini, L.; Hirsch, E. PI3K Inhibition in Inflammation: Toward Tailored Therapies for Specific Diseases. *BioEssays* **2010**, *32*, 185–196.

(51) Costa, C.; Martin-Conte, E. L.; Hirsch, E. Phosphoinositide 3-Kinase p110γ in Immunity. *IUBMB Life* **2011**, *63*, 707–713.

(52) Fung-Leung, W. P. Phosphoinositide 3-Kinase delta (PI3K δ) in Leukocyte Signaling and Function. *Cell. Signalling* **2011**, *23*, 603–608.

(53) Lee, K. S.; Lee, H. K.; Hayflick, J. S.; Lee, Y. C.; Puri, K. D. Inhibition of Phosphoinositide 3-Kinase delta Attenuates Allergic Airway Inflammation and Hyperresponsiveness in Murine Asthma Model. *FASEB J.* **2006**, *20*, 455–465.

(54) Shimada, M.; Murata, T.; Fuchikami, K.; Tsujishita, H.; Omori, N.; Kato, I.; Miura, M.; Urbahns, K.; Gantner, F.; Bacon, K. Preparation of Fused Azole-pyrimidine Derivatives as PI3K Inhibitors with Therapeutic Uses. WO2004029055A1, 2004.

(55) Hayakawa, M.; Kaizawa, H.; Moritomo, H.; Koizumi, T.; Ohishi, T.; Yamano, M.; Okada, M.; Ohta, M.; Tsukamoto, S.; Raynaud, F. I.; Workman, P.; Waterfield, M. D.; Parker, P. Synthesis and Biological Evaluation of Pyrido[3',2':4,5]furo[3,2-d]pyrimidine Derivatives as Novel PI3 Kinase P110 α Inhibitors. *Bioorg. Med. Chem. Lett.* **2007**, *17*, 2438–2442.

(56) Barber, D. F.; Bartolomé, A.; Hernandez, C.; Flores, J. M.; Fernandez-Arias, C.; Rodríguez-Borlado, L.; Hirsch, E.; Wymann, M.; Balomenos, D.; Carrera, A. C. Class I_B-Phospatidylinositol 3-Kinase (PI3K) Deficiency Ameliorates I_A- PI3K-Induced Systemic Lupus but Not T Cell Invasion. *J. Immunol.* **2006**, *176*, 589–593.

(57) Martin, A. L.; Schwartz, M. D.; Jameson, S. C.; Shimizu, Y. Selective Regulation of CD8 Effector T Cell Migration by the $p110\gamma$

Isoform of Phophatidylinositol 3-Kinase. J. Immunol. 2008, 180, 2081–2088.

(58) Patton, D. T.; Garden, O. A.; Pearce, W. P.; Clough, L. E.; Monk, C. R.; Leung, E.; Rowan, W. C.; Sancho, S.; Walker, L. S. K.; Vanhaesebroeck, B.; Okkenhaug, K. Cutting Edge: The Phosphoinositide 3-Kinase p110 δ Is Critical for the Function of CD4+CD25+Foxp3+ Regulatory T Cells. *J. Immunol.* **2006**, *177*, 6598–6602.

(59) Zhang, T. T.; Okkenhaug, K.; Nashed, B. F.; Puri, K. D.; Knight, Z. A.; Shokat, K. M.; Vanhaesebroeck, B.; Marshall, A. J. Genetic or Blockade of P110 δ Phosphoinositide 3-Kinase Enhances IgE Production. J. Allergy Clin. Immun. **2008**, 122 (4), 811–819.

(60) Park, S. J.; Lee, K. S.; Kim, S. R.; Min, K. H.; Moon, H.; Lee, M. H.; Chung, C. R.; Han, H. J.; Puri, K. D.; Lee, Y. C. Phosphoinositide 3-Kinase δ Inhibitor Suppresses Interleukin-17 Expression in a Murine Asthma Model. *Eur. Respir. J.* **2010**, *36*, 1448–1459.

(61) Marwick, J. A.; Caramori, G.; Stevenson, C. S.; Casolari, P.; Jazrawi, E.; Barnes, P. J.; Ito, K.; Adcock, I. M.; Kirkham, P. A.; Papi, A. Inhibition of PI3K δ Restores Glucocorticoid Function in Smoking Induced Airway Inflammation in Mice. *Am. J. Respir. Crit. Care Med.* **2009**, *179*, 542–548.

(62) Wei, X.; Han, J.; Chen, Z. Z.; Qi, B.-W.; Wang, G.-C.; Ma, Y.-H.; Zheng, H.; Luo, Y.-F.; Wei, Y.-Q.; Chen, L.-J. A Phosphoinositide 3-Kinase-γ Inhibitor AS605240 Prevents Bleomycin-Induced Pulmonary Fibrosis in Rats. *Biochem. Biophys. Res. Commun.* **2010**, 397, 311–317.

(63) van Dop, W. A.; Marengo, S.; te Velde, A. A.; Ciraolo, E.; Franco, I.; ten Kate, F. J.; Boeckxstaens, G. E.; Hardwick, J. C.; Hommes, D. W.; Hirsch, E.; van den Brink, G. R. The Absence of Functional PI3K γ Prevents Leukocyte Recruitment and Ameliorates DSS-Induced Colitis in Mice. *Immunol. Lett.* **2010**, *131*, 33–39.

(64) González-García, A.; Sánchez-Ruiz, J.; Flores, J. M.; Carrera, A. C. Phosphatidylinositol 3-Kinase γ Inhibition Ameliorates Inflammation and Tumor Growth in a Model of Colitis-Associated Cancer. *Gastroenterology* **2010**, *138*, 1374–1383.

(65) Gruen, M.; Rose, C.; König, C.; Gajda, M.; Wetzker, R.; Bräuer, R. Loss of Phosphoinositide 3-Kinase γ Decreases Migration and Activation of Phagocytes but Not T-Cell Activation in Antigen-Induced Arthritis. *BMC Musculoskelatal Disord.* **2010**, *11*, 1–11.

(66) Puri, K. D.; Steiner, B. H.; Kashishian, A. S.; Chen, H.; Gallatin, W. M.; Giese, N. A. IC87114, a Selective Inhibitor of PI3K δ Suppresses Joint Inflammation and Bone Erosion in Collagen-Induced Arthritis in Rat. J. Immunol. **2009**, 182 (Suppl. 50), 14.

(67) Boyle, D. L.; Rommel, C.; Topolewski, K.; Firestein, G. S. A Novel PI3Kinase/Inhibitor Suppresses Collagen-Induced Arthritis. *Arthritis Rheum.* **2009**, *60* (Suppl. 10), 669.

(68) Hayer, S.; Pundt, N.; Peters, M. A.; Wunrau, C.; Kühnel, I.; Neugebauer, K.; Strietholt, S.; Zwerina, J.; Korb, A.; Penninger, J.; Joosten, L. A. B.; Gay, S.; Rückle, T.; Schett, G.; Pap, T. PI3Ky Regulates Cartilage Damage in Chronic Inflammatory Arthritis. *FASEB J.* **2009**, *23*, 4288–4298.

(69) Doukas, J.; Wrasidlo, W.; Noronha, G.; Dneprovskaia, E.; Fine, R.; Weis, S.; Hood, J.; Demaria, A.; Soll, R.; Cheresh, D. Phosphoinositide 3-Kinase γ/δ Inhibition Limits Infarct Size after Myocardial Ischemia/Reperfusion Injury. *Proc. Natl. Acad. Sci. U.S.A.* **2006**, 103, 19866–19871.

(70) Siragusa, M.; Katare, R.; Meloni, M.; Damilano, F.; Hirsch, E.; Emanueli, C.; Madeddu, P. Involvement of Phosphoinositide 3-Kinase γ in Angiogenesis and Healing of Experimental Myocardial Infarction in Mice. *Circ. Res.* **2010**, *106*, 757–768.

(71) Rodrigues, D. H.; de Carvalho Vilela, M.; da Silva Bacelos, L.; Pinho, V.; Teixeira, M. M.; Teixeira, A. L. Absence of PI3Kγ Leads to Increased Leukocyte Apoptosis and Diminished Severity of Experimental Autoimmune Encephalomyelitis. *J. Neuroimmunol.* **2010**, *222*, 90–94.

(72) Barber, D. F.; Bartolomé, A.; Hernandez, C.; Flores, J. M.; Redondo, C.; Fernandez-Arias, C.; Camps, M.; Rückle, T.; Schwaarz, M. K.; Rodríguez, S.; Martinez-A, C.; Balomenos, D.; Rommel, C.; Carrera, A. C. PI3Ky Inhibition Blocks Glomerulonephritis and Extends Lifespan in a Mouse Model of Systemic Lupus. Nat. Med. 2005, 11, 933-935.

(73) Swat, W.; Montgrain, V.; Doggett, T. A.; Douangpanya, J.; Puri, K.; Vermi, W.; Diacovo, T. G. Essential Role of PI3K δ and PI3K γ in Thymocyte Survival. *Blood* **2006**, *107*, 2415–2422.

(74) Randis, T. M.; Puri, K. D.; Zhou, H.; Diacovo, T. G. Role of PI3K δ and PI3K γ in Inflammatory Arthritis and Tissue Localization of Neutrophils. *Eur. J. Immunol.* **2008**, *38*, 1215–1224.

(75) Rommel, C. Taking PI3K δ and PI3K γ One Step Ahead: Dual Active PI3K δ/γ Inhibitors for the Treatment of Immune-Mediated Inflammatory Diseases. In *Phosphoinositide 3-Kinase in Health and Disease*; Rommel, C., Vanhaesebroeck, B., Vogt, P. K., Eds.; Current Topics in Microbiology and Immunology; Springer: New York, 2010; Vol. 1, pp 279–301.

(76) Chuckowree, I.; Folkes, A.; Hancox, T.; Shuttleworth, S. Preparation of Substituted Thienopyrimidine as Inhibitor of Phosphatidylinositol 3-Kinase. WO2007129161A2, 2007.

(77) National Institutes of Health. Identifier: NCT01493843; NCT01437566. ClinicalTrials.gov.

(78) Folkes, A. J.; Ahmadi, K.; Alderton, W. K.; Alix, S.; Baker, S. J.; Box, G.; Chuckowree, I. S.; Clarke, P. A.; Depledge, P.; Eccles, S. A. E.; Friedman, L. S.; Hayes, A.; Hancox, T. C.; Kugendradas, A.; Lensun, L.; Moore, P.; Olivero, A. G.; Pang, J.; Patel, S.; Pergl-Wilson, G. H.; Raynaud, F. I.; Robson, A.; Saghir, N.; Salphati, L.; Sohal, S.; Ultsch, M. H.; Valenti, M.; Wallweber, H. J.; Wan, N. C.; Wiesmann, C.; Workman, P.; Zhyvoloup, A.; Zvelebil, M. J.; Shuttleworth, S. J. The Identification of 2-(1H-Indazol-4-yl)-6-(4-methanesulfonyl-piperazin-1-ylmethyl)-4-morpholin-4-yl-thieno[3,2-*d*]pyrimidine (GDC-0941) as a Potent, Selective, Orally Bioavailable Inhibitor of Class I PI3 Kinase for the Treatment of Cancer. *J. Med. Chem.* **2008**, *51*, 5522– 5532.

(79) Baker, S. J.; Goldsmith, P. J.; Hancox, T. C.; Pegg, N. A.; Price, S.; Shuttleworth, S. J.; Sohal, S. Preparation of Thienopyrimidine Derivatives as PI3 Kinase Inhibitors. WO2007122410A1, 2007.

(80) Hancox, T. C.; Pegg, N. A.; Nadin, A. J.; Price, S. Preparation of Thienopyrimidine Derivatives as Inhibitors of PI3 Kinase. WO2009053715A1, 2009.

(81) Shuttleworth, S. J.; Folkes, A. J.; Chuckowree, I. S.; Wan, N. C.; Hancox, T. C.; Baker, S. J.; Sohal, S.; Latif, M. A. Preparation of Thienopyrimidines as Phosphatidylinositol 3 Kinase Inhibitors. WO2006046035A1, 2006.

(82) Castanedo, G.; Goldstein, D.; Kondru, R. K.; Lucas, M.; Palmer, W.; Price, S.; Safina, B.; Savy, P. P. A.; Seward, E. M.; Sutherlin, D. P.; Sweeney, Z. K. Bicyclic Indole-pyrimidine PI3K Inhibitor Compounds Selective for P110 delta, and Methods of Use. U.S. Pat. Appl. 20100305084A1, 2010.

(83) Li, J.; Safina, B.; Sutherlin, D. P.; Sweeney, Z. Preparation and Methods of Use of Purine Compounds Selective for PI3 Kinase p110 delta Isoform. WO2012007493A1, 2012.

(84) Castanedo, G.; Chan, B.; Lucas, M. C.; Safina, B.; Sutherlin, D. P.; Sweeney, Z. K. Preparation of Pyrido[3,2-d]pyrimidine as PI3K δ Inhibitors. WO2011101429A1, 2011.

(85) Castanedo, G.; Chan, B.; Goldstein, D. M.; Kondru, R. K.; Lucas, M. C.; Palmer, W. S.; Price, S.; Safina, B.; Savy, P. P. A.; Seward, E. M.; Sutherlin, D. P.; Sweeney, Z. K. Preparation of Bicyclic Pyrimidines as Selective Inhibitors of the p110 delta Isoform of PI3K for Treating Inflammation, Immune Diseases and Cancers. WO2010138589A1, 2010.

(86) Bruce, I.; Weitz, S. G. Preparation of (Thiazolylureido)propionamides as Inhibitors of PI3K delta Enzyme. WO2008000421A2, 2008.

(87) White, S. L. Thienopyrimidinones as δ -PI3K Inhibitors and Their Preparation, Pharmaceutical Compositions and Use in the Treatment of Inflammatory Disorders and Cancers. WO2008064018A1, 2008.

(88) White, S. L.; Ruan, F.; Kesicki, E. A.; Thorsett, E.; Farouz, F. Preparation of Purine Pyrimidinone Compounds as Inhibitors of Human Phosphatidyl-Inositol 3-Kinase delta for Treating Immune, Inflammatory, and Other Diseases. WO2009064802A2, 2009.

(89) Chen, Y.; Cushing, T. D.; Hao, X.; He, X.; Reichelt, A.; Rzasa, R. M.; Seganish, J.; Shin, Y.; Zhang, D. Heterocyclic Compounds and Their Uses in the Treatment of p110 δ -Mediated Diseases and Their Preparation. WO2008118454A2, 2008.

(90) Chen, Y.; Cushing, T. D.; Duquette, J. A.; Gonzalez, L. d. T. F.; Hao, X.; He, X.; Lucas, B.; McGee, L. R.; Reichelt, A.; Rzasa, R. M.; Seganish, J.; Shin, Y.; Zhang, D. Heterocyclic Compounds and Their Uses in the Treatment of p110 δ -Mediated Diseases and Their Preparation. WO2008118468A1, 2008.

(91) Cushing, T. D.; Dransfield, P. J.; Gonzalez, L. d. T. F.; Johnson, M. G.; Kohn, T. J.; Pattaropong, V.; Simard, J. L. Preparation of Heteroaryl Compounds as Inhibitors of PI3K Inhibitors. WO2010151791A1, 2010.

(92) Brown, M.; Chen, Y.; Cushing, T. D.; Gonzalez, L. d. T. F.; He, X.; Kohn, T. J.; Lohman, J. W.; Pattaropong, V.; Seganish, J.; Shin, Y.; Simard, J. L. Preparation of Heterocyclic Compounds as Therapeutic PI3K δ Inhibitors. WO2010151737A2, 2010.

(93) Bui, M. H. T.; Chen, Y.; Cushing, T. D.; Duquette, J. A.; Fisher, B.; Gonzalez, L. d. T. F.; Hao, X.; He, X.; Johnson, M. G.; Lucas, B. Preparation of Quinolinylethylamino-pyrimidinecarbonitrile Derivative as Inhibitors of PI3K. WO2010151740A2, 2010.

(94) Cushing, T. D.; Hao, X.; Lohman, J. W.; Shin, Y. Preparation of Pyrido[1,2-*a*]pyrimidine and Quinoline Derivatives as Inhibitors of PI3K. WO2010151735A2, 2010.

(95) Chen, Y.; Cushing, T. D.; Hao, X.; He, X.; Reichelt, A.; Rzasa, R. M.; Seganish, J.; Shin, Y.; Zhang, D. δ 3-Substituted Quinoline or Quinoxaline Derivatives and Their Use as Phosphatidylinositol 3-kinase (PI3K) Inhibitors and Their Preparation. WO2008118455A1, 2008.

(96) Cushing, T. D.; Duquette, J. A.; He, X.; Lohman, J. W.; Shin, Y. Preparation of Substituted Bicyclic Heteroaryl Compounds as PI3K Kinase Inhibitors. WO2011123751A2, 2011.

(97) Lucas, B. Preparation of Thienopyridinylethylpurinamine Derivatives for Use as PI3K Activity Inhibitors. WO2012003262A1, 2012.

(98) Cushing, T. D.; Gonzalez, L. d. T. F.; Hao, X.; Shin, Y. Heterocyclic Compounds as PI3K Inhibitors and Their Preparation and Use in the Treatment of Diseases. WO2011075628A1, 2011.

(99) Duquette, J. A.; Lucas, B. Preparation of Quinoline Derivatives for Use as PI3K Inhibitors. WO2012003271A1, 2012.

(100) Dransfield, P. J. Preparation of Quinoline Derivatives for Use as PI3K Inhibitors. WO2012003278A1, 2012.

(101) Dransfield, P. J.; Gonzalez, L. d. T. F.; Kohn, T. J.; Pattaropong, V.; Simard, J. L. Preparation of Quinoline Derivatives for Use as PI3K Inhibitors. WO2012003283A1, 2012.

(102) National Institutes of Health. Identifier: NCT01300026. ClinicalTrials.gov.

(103) Intellikine Joins the Takeda Family of Companies. http:// www.intellikine.com/.

(104) Ren, P.; Liu, Y.; Wilson, T. E. Antitumor PI3 Kinase Modulators. WO2009046448A1, 2009.

(105) Ren, P.; Liu, Y.; Wilson, T. E.; Chan, K.; Rommel, C.; Li, L. Preparation of Pyrazolo Pyrimidines and Purine Compounds as Therapeutic PI3 Kinase Modulators. WO2009088986A1, 2009.

(106) Ren, P.; Liu, Y.; Wilson, T. E.; Li, L. Heterocyclic Compounds and Uses Thereof as Phosphatidylinositol 3'-Kinase Modulators. WO2010129816A2, 2010.

(107) Ren, P.; Liu, Y.; Li, L.; Chan, K.; Wilson, T. E. Preparation of Benzothiazoles as Protein and Lipid Kinase Inhibitors. WO2009114874A2, 2009.

(108) Ren, P.; Liu, Y.; Li, L.; Chan, K.; Wilson, T. E.; Martin, M.; Rommel, C. Preparation of 1*H*-Pyrazolo[3,4-*d*]pyrimidine Compounds as Kinase Inhibitors and Methods of Use as Therapeutic Agents. WO2010006086A2, 2010.

(109) Bartok, B.; Boyle, D. L.; Liu, Y.; Ren, P.; Ball, S. T.; Bugbee, W. D.; Rommel, C.; Firestein, G. S. PI3 Kinase δ Is a Key Regulator of Synoviocyte Function in Rheumatoid Arthritis. *Am. J. Pathol.* **2012**, *180*, 1906–1916.

(110) Knight, Z. A.; Shokat, K. M.; Williams, O. Preparation of Pyrazolopyrimidinylmethyl Quinazolinones as PI3 Kinase Antagonists. WO2008127226A2, 2008.

(111) Berndt, A.; Miller, S.; Williams, O.; Le, D. D.; Houseman, B. T.; Pacold, J. I.; Gorrec, F.; Hon, W.-C.; Liu, Y.; Rommel, C.; Gaillard, P.; Rückle, T.; Schwarz, M. K.; Shokat, K. M.; Shaw, J. P.; Williams, R. L. The P110 δ Crystal Structure: Mechanisms for Selectivity and Potency of New PI(3)K Inhibitors. *Nat. Chem. Biol.* **2010**, *6*, 117–124.

(112) Allen, D. R.; Buckley, G. M.; Buerli, R.; Davenport, R. J.; Kinsella, N.; Lock, C. J.; Lowe, C.; Mack, S. R.; Pitt, W. R.; Ratcliffe, A. J.; Richard, M. D.; Sabin, V. M.; Sharpe, A.; Tait, L. J.; Warrelow, G. J.; Williams, S. C. Preparation of Quinoxaline and Quinoline Derivatives as Kinase Inhibitors. WO2009081105A2, 2009.

(113) Beevers, R. E.; Mack, S. R.; Sabin, V. M. Preparation of Quinoline Derivatives for Use as PI3 Kinase Inhibitors. WO2010061180A1, 2010.

(114) Allen, D. R.; Brown, J. A.; Buerli, R.; Haughan, A. F.; MacDonald, J. D.; Matteucci, M.; Owens, A. P.; Raphy, G.; Saville-Stones, E. A.; Sharpe, A. Fused Pyridines and Related Compounds as Kinase Inhibitors and Their Preparation and Use for the Treatment of Diseases. WO2010092340A1, 2010.

(115) Allen, D. R.; Brown, J. A.; Buerli, R.; Haughan, A. F.; Langham, B. J.; Matteucci, M.; Owens, A. P.; Raphy, G.; Sharpe, A. Preparation of Fused Bicyclic Pyrazole Derivatives as PI3 Kinase Inhibitors. WO2010133836A1, 2010.

(116) Rewcastle, G. W.; Shepherd, P. R.; Chaussade, C.; Denny, W. A.; Gamage, S. A. Substituted Pyrimidines and Triazines as Inhibitors of PI3K Enzymes Useful in the Treatment of Cancer and Preparation and Pharmaceutical Compositions Thereof. WO2009120094A2, 2009. (117) Kawashima, S.; Matsuno, T.; Yaguchi, S.; Sasahara, H.; Watanabe, T. Preparation of Heterocyclic Compounds as Antitumor Agents. WO2002088112A1, 2002.

(118) Yaguchi, S.; Fukui, Y.; Koshimizu, I.; Yoshimi, H.; Matsuno, T.; Gouda, H.; Hirono, S.; Yamazaki, K.; Yamori, T. Antitumor Activity of ZSTK474, a New Phosphatidylinositol 3-Kinase Inhibitor. *J. Natl. Cancer Inst.* **2006**, *98*, 545–556.

(119) Kong, D.; Yamori, T. ZSTK474 is an ATP-Competitive Inhibitor of Class 1 Phosphatidylinositol 3-Kinase Isoforms. *Cancer Sci.* **2007**, *98*, 1638–1642.

(120) Toyama, S.; Tamura, N.; Haruta, K.; Karakida, T.; Mori, S.; Watanabe, T.; Yamori, T.; Takasaki, Y. Inhibitory Effects of ZSTK474, a Novel Phosphoinositide 3-Kinase Inhibitor, on Osteoclasts and Collagen-Induced Arthritis in Mice. *Arthritis Res. Ther.* **2010**, *12* (R92), 1–11.

(121) Liang, C.; Li, Z.-G. Preparation of PI3K Isoform Selective Inhibitors. WO2010005558A2, 2010.

(122) Ivashchenko, A. V.; Tkachenko, S. E.; Okun, I. Y. M.; Savchuk, N. F.; Ivashchenko, A. A.; Kravchenko, D. V.; Shmidt-Kittler, O.; Rago, K.; Papadopoulos, N.; Velcuelsu, V.; Vogelstein, B.; Kinzler, K. W.; Trifilenkov, A. S.; Rizhova, E. A.; Jiuxiang, Z. Annelated Azahetero-cycles Comprising Pyrimidine Fragment, Method for the Production Thereof and (PI3Ks) Kinase Inhibitors. WO 2009011617A2, 2009.

(123) Gaillard, P.; Quattropani, A.; Pomel, V.; Rueckle, T.; Klicic, J.; Church, D. Preparation of Pyrazine Derivatives, Particularly N-[3-(Oxyphenylamino)quinoxalin-2-yl]sulfonamides, as PI3K Inhibitors. WO2007023186A1, 2007.

(124) Ren, P.; Liu, Y.; Li, L.; Chan, K.; Wilson, T. E. Preparation of Quinolinylmethylpyrazolopyrimidine Derivatives and Analogs for Use as Kinase Inhibitors. WO2010036380A1, 2010.

(125) Takahashi, F.; Imada, S.; Shiwaku, M.; Kato, K.; Fukahori, H. Preparation of 2-(Difluoromethyl)-1-(morpholinoheterocyclyl)benzimidazole Derivatives as Selective Phosphatidyl Inositol-3-kinase (PI3K δ) Inhibitors. WO2010092962A1, 2010.

(126) Shuttleworth, S. J.; Silva, F. A.; Cecil, A. R. L.; Hill, T. J. Preparation of Benzo[1,3]oxazinone Derivatives as PI3 Kinase Inhibitors. WO2011012883A1, 2011.

(127) Shuttleworth, S. J.; Silva, F. A. Preparation of Naphthyridine Derivatives as PI3K Inhibitors for the Treatment of Cancer and Immune-Inflammatory Diseases. WO2011135351A1, 2011.

(128) Karus Therapeutics. http://www.karustherapeutics.com/.

(129) Baldwin, I. R.; Down, K. D.; Faulder, P.; Gaines, S.; Hamblin, J. N.; Jones, K. L.; Le, J.; Lunniss, C. J.; Parr, N. J.; Ritchie, T. J.; Smethurst, C. A. P.; Washio, Y. Preparation of 4-Carboxamide Indazole Derivatives Useful as Inhibitors of PI3-Kinases. WO2009147187A1, 2009.

(130) Hamblin, J. N.; Jones, P. S.; Keeling, S. E.; Le, J.; Mitchell, C. J.; Parr, N. J. Preparation of Oxazole-Substituted Indazole Derivatives for Use as PI3-Kinase Inhibitors. WO2010125082A1, 2010.

(131) Baldwin, I. R.; Down, K. D.; Faulder, P.; Gaines, S.; Hamblin, J. N.; Le, J.; Lunniss, C. J.; Parr, N. J.; Ritchie, T. J.; Robinson, J. E.; Simpson, J. K.; Smethurst, C. A. P. Preparation of *N*-Indazolyl Carboxamides as PI3-Kinase Inhibitors. WO2011067364A1, 2011.

(132) Hamblin, J. N.; Jones, P. S.; Keeling, S. E.; Le, J.; Mitchell, C. J.; Parr, N. J.; Willacy, R. D. Preparation of Polymorphs and Salts of *cis-N*-[5-[4-[5-[[2,6-Dimethyl-4-morpholinyl]methyl]-2-oxazolyl]-1*H*-indazol-6-yl]-2-methoxy-3-pyridinyl]-methanesulfonamide as PI3K δ Inhibitors. WO2012032067A1, 2012.

(133) National Institutes of Health. Identifier: NCT01462617. ClinicalTrials.gov.

(134) Muthuppalaniappan, M.; Viswanadha, S.; Babu, G.; Vakkalanka, S. K. V. S. Preparation of Chromen-4-one Compounds as Therapeutic PI3K Kinase Modulators. WO2011055215A2, 2011.

(135) King-Underwood, J.; Ito, K.; Murray, P. J.; Hardy, G.; Brookfield, F. A.; Brown, C. J. Preparation of Pyrazolopyrimidine Compounds as Phosphoinositide 3-Kinase (PI3 Kinase) Inhibitors. WO2011048111A1, 2011.

(136) Li, Y.-L.; Metcalf, B. W.; Combs, A. P. Preparation of Pyrimidinones as PI3K Inhibitors. WO2011008487A1, 2011.

(137) Combs, A. P.; Li, Y.-L.; Yue, E. W.; Sparks, R. B. Preparation of Substituted Heteroaryl Fused Derivatives as PI3K Inhibitors. WO2011075643A1, 2011.

(138) Combs, A. P.; Sparks, R. B.; Yue, E. W. Preparation of Imidazopyridinylethylpurinamine Derivatives and Analogs for Use as PI3K Inhibitors. WO2011130342A1, 2011.

(139) (a) Li, Y.-L.; Combs, A. P. Fused Pyrrole Derivatives as PI3K Inhibitors U.S. Pat. Appl. 20110312979A1, 2011. (b) Li, Y.-L.; Combs, A. P.; Yue, E. W.; Li, H.-Y. Preparation of Substituted Fused Aryl and Heteroaryl Derivatives as PI3K Inhibitors. WO2011075630A1, 2011. (140) Pathway Therapeutics. http://www.pathwaytx.com/.

(141) Mattheakis, L.; Kearney, P.; Jaegar, C.; Plonowski, A.; Zhang, J.; Yu, P. Presented at the 22nd EORTC-NCI-AACR Symposium on Molecular Targets and Cancer Therapeutics, Berlin, Germany, November 16–19, 2010; Abstract 219.

(142) Kearney, P. Preparation of Heterocyclyl-Substituted Purine Derivatives as Inhibitors of PI3K-delta for the Treatment of Cancer. WO2012037226A1, 2012.

(143) Leahy, J. W. Preparation of Heterocyclyl-Substituted Purine Derivatives as Inhibitors of PI3K-delta for the Treatment of Cancer. WO2012037204A1, 2012.

(144) Fowler, K. W.; Huang, D.; Kesicki, E. A.; Ooi, H. C.; Oliver, A. R.; Ruan, F.; Treiberg, J. Preparation of Purine Quinazolinones as Inhibitors of Human Phosphatidylinositol 3-Kinase delta. WO2005113556A1, 2005.

(145) Ikeda, H.; Hideshima, T.; Fulciniti, M.; Perrone, G.; Miura, N.; Yasui, H.; Okawa, Y.; Kiziltepe, T.; Santo, L.; Vallet, S.; Cristea, D.; Calabreseqq, E.; Gorgun, G.; Raje, N. S.; Richardson, P.; Munshi, N. C.; Lannutti, B. J.; Puri, K. D.; Giese, N. A.; Anderson, K. C. PI3K/ p110 δ Is a Novel Therapeutic Target in Multiple Myeloma. *Blood* **2010**, *116*, 1460–1468.

(146) Gallatin, M. W.; Ulrich, R. G.; Giese, N. Use of Quinazoline Derivatives for the Treatment of Hematologic Malignancies, Inflammatory and Autoimmune Disorders. WO2010057048A1, 2010.

(147) Flinn, I. W.; Byrd, J. C.; Furman, R. R.; Brown, J. R.; Benson, D. M.; Coutre, S. E.; Kahl, B. S.; Smith, B. D.; Wagner-Johnston, N. D.; Spurgeon, S. E.; Giese, N. A.; Yu, A. S. Presented at the American Society of Hematology Annual Meeting, New Orleans, LA, U.S., 2009; Abstract 922.

(148) Webb, H. K; Chen, H.; Yu, A. S.; Peterman, S.; Holes, L.; Lannutti, B.; Miller, L. L.; Ulrich, R. G. Presented at the American Society of Hematology Annual Meeting, Orlando, FL, U.S., 2010; Abstract 1774.

(149) National Institutes of Health. Identifier: NCT00836914, ClinicalTrials.gov.

(150) National Institutes of Health. Identifier: NCT01066611. ClinicalTrials.gov.

(151) Ulrich, R. G. Treatment of Liver Disorders with PI3K Inhibitors. WO2011011550A1, 2011.

(152) Sadhu, C.; Dick, K.; Tino, W. T.; Staunton, D. E. Selective Role of PI3K δ in Neutrophil Inflammatory Responses. *Biochem. Biophys. Res. Commun.* **2003**, 308, 764–769.

(153) Quattropani, A.; Dorbais, J.; Covini, D.; Desforges, G.; Rueckle, T. Thiazole Derivatives and Use Thereof. U.S. Pat. Appl. 20080188531A1, 2008.

(154) Barvian, N. C.; Kolz, C. N.; Para, K. S.; Patt, W. C.; Visnick, M. Preparation of Benzoxazin-3-ones and Derivatives as Inhibitors of PI3K Kinase for Treating Inflammations, Cardiovascular Diseases and Cancers. WO2004052373A1, 2004.

(155) Connolly, M.; Gogliotti, R. D.; Lee, H. T.; Plummer, M. S.; Sexton, K. E.; Visnick, M. Preparation of N-Tetrazolyl Benzo[b]thiophenecarboxamides as Phosphoinositide-3-kinase (PI3K) Inhibitors for the Treatment of Cancer, Inflammatory and Cardiovascular Diseases. WO2004108713A1, 2004.

(156) Connolly, M. K.; Gogliotti, R. D.; Plummer, M. S.; Visnick, M. Preparation of Morpholinyl-Pyrimidine Derivatives as Inhibitors of Phosphoinositide-3-kinases. WO2005042519A1, 2005.

(157) Wilson, F.; Ramsden, N.; Bell, K.; Cansfield, A.; Burckhardt, S.; Taylor, J.; Sunose, M.; Middlemiss, D. Preparation of (Aryltriazolopyridinyl)amine Derivatives for Use as Kinase Inhibitors. WO2008025821A1, 2008.

(158) Wang, T.; Green, J.; Cornebise, M.; Ledford, B.; Parsons, J.; Tanner, A.; Westcott, J. Preparation of Thiazolyl Furan-2-carboxylate Derivatives as Inhibitors of Phosphatidylinositol 3-Kinase. WO2008027584A2, 2008.

(159) Aronov, A.; Bandarage, U. K.; Cottrell, K.; Davies, R.; Krueger, E.; Ledeboer, M.; Ledford, B.; Le, T. A.; Liao, Y.; Messersmith, D.; Wang, T.; Xu, J. Preparation of Tetrahydrothiazolopyridine Derivatives for Use as Phosphatidylinositol 3-Kinase Inhibitors. WO2010096389A1, 2010.

(160) Wang, T.; Aronov, A.; Cornebise, M.; Maltais, F.; Ledoboer, M.; Le, T. A.; Marone, V.; Messersmith, D.; Cottrell, K. Pyridinylbenzothiazole-urea Derivatives as Phosphatidylinositol 3-Kinase Inhibitors and Their Preparation, Pharmaceutical Compositions and Use in the Treatment of Diseases. WO2009129211A1, 2009.

(161) Aronov, A.; Come, J.; Cottrell, K.; Grey, R.; LeTiran, A.; Marone, V.; Martinez-Botella, G.; Messersmith, D.; Huck, E. P. Preparation of Benzothiazole Derivatives for Use as PI3K Inhibitors and Useful in the Treatment of Autoimmune Diseases. WO2010135014A1, 2010.

(162) Bruce, I.; Hayler, J. F.; Bloomfield, G. C.; Edwards, L.; Cox, B.; Howsham, C. Preparation of Pyrazolo[3,4-*d*]pyrimidine Derivatives for Treating Respiratory Disorders. WO2007134828A1, 2007.

(163) Bloomfield, G. C.; Bruce, I.; Hayler, J. F.; Leblanc, C.; Le, G. D. M.; McCarthy, C. Preparation of Phenylthiazolylureas as Inhibitors of Phosphatidylinositol 3-Kinase. WO2005021519A2, 2005.

(164) Bruce, I.; Budd, E.; Edwards, L.; Howsham, C. Preparation of Pyrazine Compounds as Inhibitors of Phosphatidylinositol 3-Kinase Enzymes in the Treatment of Inflammatory and Allergic Conditions. WO2009115517A2, 2009.

(165) Moffat, D. C. F.; Davies, S.; Alesso, S. M.; Launay, D. F. M. Preparation of 4-Methyl-5-(alkylsulfonylphenyl)thiazoles, Particularly Amino Acid Derivatives of Thiazoles, as Inhibitors of PI3 Kinase Useful for Treating Autoimmune, Inflammatory and Proliferative Diseases. WO2007129048A1, 2007.

(166) Grauert, M.; Maier, U.; Hoffmann, M.; Scheuerer, S.; Joergensen, A. T.; Pautsch, A.; Brandl, T.; Hoenke, C.; Breitfelder,

S.; Erb, K.; Pieper, M.; Pragst, I. Thiazolyl-dihydro-indazoles. U.S. Pat. Appl. 20090093474A1, 2009.

(167) Rueckle, T.; Quattropani, A.; Pomel, V.; Dorbais, J.; Covini, D.; Bischoff, A. Preparation of Pyridine Methylene Thioxothiazolidinones as Phosphoinositide Inhibitors. WO2006024666A1, 2006.

(168) Taniyama, D.; Kano, K.; Ishibashi, K.; Endoh, T. Preparation of Pyrimidine Derivative and Pyridine Derivative Both Having Phosphatidyl Inositol-3 Kinase (PI3K) Inhibitory Activity. WO2010061903A1, 2010.

(169) Fujioka, M.; Mitsumori, S.; Kugimiya, A.; Taniyama, D. Preparation of 3,4-Dihydro-2*H*-pyrido[3,2-*b*]-1,4-oxazine Derivatives as PI3K Inhibitors for Treatment of Inflammation. WO2010027002A1, 2010.

(170) Leahy, J. W.; Buhr, C. A.; Johnson, H. W. B.; Kim, B. G.; Baik, T.; Cannoy, J.; Forsyth, T. P.; Jeong, J. W.; Lee, M. S.; Ma, S.; Noson, K.; Wang, L.; Williams, M.; Nuss, J. M.; Brooks, E.; Foster, P.; Goon, L.; Heald, N.; Holst, C.; Jaeger, C.; Lam, S.; Lougheed, J.; Nguyen, L.; Plonowski, A.; Song, J.; Stout, T.; Wu, X.; Yakes, M. F.; Yu, P.; Zhang, W.; Lamb, P.; Raeber, O. Discovery of a Novel Series of Potent and Orally Bioavailable Phosphoinositide 3-Kinase γ Inhibitors. *J. Med. Chem.* **2012**, *55*, 5467–5482.

(171) Alexander, R.; Balasundaram, A.; Batchelor, M.; Brookings, D.; Crépy, K.; Crabbe, T.; Deltent, M.-F.; Driessens, F.; Gill, A.; Harris, S.; Hutchinson, G.; Kulisa, C.; Merriman, M.; Mistry, P.; Parton, T.; Turner, J.; Whitcombe, I.; Wright, S. 4-(1,3-Thiazol-yl)morpholine Derivative as Inhibitors of Phosphoinositide 3-Kinase. *Bioorg. Med. Chem. Lett.* **2008**, *18*, 4316–4320.

(172) Perry, B.; Alexander, R.; Bennett, G.; Buckley, G.; Ceska, T.; Crabbe, T.; Dale, V.; Gowers, L.; Horsley, H.; James, L.; Jenkins, K.; Crépy, K.; Kulisa, C.; Lightfoot, H.; Lock, C.; Mack, S.; Morgan, T.; Nicolas, A.-L.; Pitt, W.; Sabin, V.; Wright, S. Achieving Multi-Isoform PI3K Inhibition in a Series of Substituted 3,4-Dihydro-2*H*-benzo-{1,4}oxazines. *Bioorg. Med. Chem. Lett.* **2008**, *18*, 4700–4704.

(173) Perry, B.; Beevers, R.; Bennett, G.; Buckley, G.; Crabbe, T.; Gowers, L.; James, L.; Jenkins, K.; Lock, C.; Sabin, V.; Wright, S. Optimization of a Series of Multi-Isoform PI3 Kinase Inhibitors. *Bioorg. Med. Chem. Lett.* **2008**, *18*, 5299–5302.

(174) Cardin, D. P.; Gaulin, J. L.; Greenspan, P. D.; Vyskocil, S.; Xu, T.; Renou, C. C. Preparation of Thiophene or Thiazole Derivatives and Their Use as PI3K Inhibitors. WO2009154741A1, 2009.

(175) Baldwin, I. R.; Down, K. D.; Faulder, P.; Gaines, S.; Hamblin, J. N.; Le, J.; Lunniss, C. J.; Parr, N. J.; Ritchie, T. J.; Simpson, J. K.; Smethurst, C. A. P. Preparation of 4-Amino-1*H*-indazole Compounds as Inhibitors of PI3-Kinases for Treatment of Asthma and COPD. WO2009147190A1, 2009.

(176) Baldwin, I. R.; Down, K. D.; Faulder, P.; Gaines, S.; Hamblin, J. N.; Jones, P. S.; Le, J.; Parr, N. J.; Ritchie, T. J.; Robinson, J. E.; Simpson, J. K.; Smethurst, C. A. P. Preparation of Indolylindazole Derivatives for Use as PI3-Kinase Inhibitors. WO2009147189A1, 2009.

(177) Hamblin, J. N.; Harrison, Z. A.; Jones, P. S.; Keeling, S. E.; Le, J.; Lunniss, C. J.; Parr, N. J. Preparation of 4-(Oxadiazol-2-yl)indazoles as Inhibitors of PI3 Kinase. WO2010102958A1, 2010.

(178) Baldwin, I. R.; Down, K. D.; Faulder, P.; Gaines, S.; Hamblin, J. N.; Le, J.; Lunniss, C. J.; Parr, N. J.; Ritchie, T. J.; Robinson, J. E.; Simpson, J. K.; Smethurst, C. A. P. Benzopyrazole Derivatives as PI3 Kinase Inhibitors and Their Preparation, Pharmaceutical Compositions and Use in the Treatment of Diseases. WO2009147188A1, 2009.

(179) Baldwin, I. R.; Down, K. D.; Faulder, P.; Gaines, S.; Hamblin, J. N.; Jones, K. L.; Jones, P. S.; Le, J.; Lunniss, C. J.; Parr, N. J.; Ritchie, T. J.; Robinson, J. E.; Simpson, J. K.; Smethurst, C. A. P. Indazole Derivatives as PI3 Kinase Inhibitors and Their Preparation and Use for the Treatment of PI3 Kinase-Mediated Diseases. WO2011067365A1, 2011.

(180) Baldwin, I. R.; Down, K. D.; Faulder, P.; Gaines, S.; Hamblin, J. N.; Jones, K. L.; Le, J.; Lunniss, C. J.; Parr, N. J.; Ritchie, T. J.; Robinson, J. E.; Smethurst, C. A. P. Indazole Derivatives as PI3 Kinase Inhibitors and Their Preparation and Use for the Treatment of PI3 Kinase-Mediated Diseases. WO2011067366A1, 2011.

(181) Williams, O.; Houseman, B. T.; Kunkel, E. J.; Aizenstein, B.; Hoffman, R.; Knight, Z. A.; Shokat, K. M. Discovery of Dual Inhibitors of the Immune Cell PI3Ks P110 δ and P110 γ : A Prototype for New Anti-Inflammatory Drugs. *Chem. Biol.* **2010**, *17*, 123–134.

(182) Ren, P.; Liu, Y.; Wilson, T. E.; Chan, K.; Rommel, C.; Li, L. Preparation of Pyrazolo Pyrimidines and Other Heterocyclic Compounds as Therapeutic P13 Kinase Modulators. WO2009088990A1, 2009.

(183) Infinity Pharmaceuticals. http://www.infi.com/.

(184) Ihle, N T.; Powis, G. Take Your PIK: Phosphatidylinositol 3-Kinase Inhibitors Race through the Clinic and toward Cancer Therapy. *Mol. Cancer Ther.* **2009**, *8*, 1–9.

(185) Walker, E. H.; Perisic, O.; Ried, C.; Stephens, L.; Williams, R. L. Structural Insights into Phosphoinositide 3-Kinase Catalysis and Signaling. *Nature* **1999**, *402*, 313–320.

(186) Walker, E. H.; Pacold, M. E.; Perisic, O.; Stephens, L.; Hawkins, P. T.; Wymann, M. P.; Williams, R. L. Structural Determinants of Phosphoinositide 3-Kinase Inhibition by Wortmannin LY294002, Quercetin, Myricetin, and Staurosporine. *Mol. Cell* **2000**, *6*, 909–919.

(187) Akritopoulou-Zanze, I.; Hajduk, P. J. Kinase-Targeted Libraries: The Design and Synthesis of Novel, Potent, and Selective Kinase Inhibitors. *Drug Discovery Today* **2009**, *14*, 291–297.

(188) Zhang, X.; Vadas, O.; Perisic, O.; Anderson, K. E.; Clark, J.; Hawkins, P. T.; Stephens, L. R.; Williams, R. L. Structure of Lipid $p110\beta/p85\beta$ Elucidates an Unusual SH2-Domain-Mediated Inhibitory Mechanism. *Mol. Cell* **2011**, *41*, 567–578.

(189) Hon, W. C.; Berndt, A.; Williams, R. L. Regulation of Lipid Binding Underlies the Activation Mechanism of Class IA PI3-Kinases. *Oncogene* **2012**, *31*, 3655–3666.

(190) Huang, C.-H.; Mandelker, D.; Schmidt-Kittler, O.; Samuels, Y.; Velculescu, V. E.; Kinzler, K. W.; Vogelstein, B.; Gabelli, S. B.; Amzel, L. M. The Structure of a Human p110 α /p85 α Complex Elucidates the Effects of Oncogenic PI3K α Mutations. *Science* **2007**, *318*, 1744–1748.

(191) Mandelker, D.; Gabelli, S. B.; Schmidt-Kittler, O.; Zhu, J.; Cheong, I.; Huang, C.-H.; Kinzler, K. W.; Vogelstein, B.; Amzel, L. M. A Frequent Kinase Domain Mutation That Changes the Interaction between PI3K α and the Membrane. *Proc. Natl. Acad. Sci. U.S.A.* **2009**, *106*, 16996–17001.

(192) Amzel, L. M.; Huang, C.-H.; Mandelker, D.; Lengauer, C.; Gabelli, S. B.; Vogelstein, B. Structural Comparisons of Class I Phosphoinositide 3-Kinases. *Nat. Rev. Cancer* **2008**, *8*, 665–669.

(193) Kuang, R.-R.; Qian, F.; Li, Z.; Wei, D.-Z. Study on Improving the Selectivity of Compounds That Inhibit Two PI3Ks (gamma and delta). *J. Mol. Model.* **2006**, *12*, 445–452.

(194) Zvelebil, M. J.; Waterfield, M. D.; Shuttleworth, S. J. Structural Analysis of PI3-Kinase Isoforms: Identification of Residues Enabling Selective Inhibition by Small Molecule ATP-Competitive Inhibitors. *Arch. Biochem. Biophys.* **2008**, 477, 404–410.

(195) Sinnamon, R. H.; McDevitt, P.; Pietrak, B. L.; Leydon, V. R.; Xue, Y.; Lehr, R.; Qi, H.; Burns, M.; Elkins, P.; Ward, P.; Vincentini, G.; Fisher, D.; Grimes, M.; Brandt, M.; Auger, K. R.; Ho, T.; Johanson, K.; Jones, C. S.; Schwartz, B.; Sweitzer, T.,D.; Kirkpatrick, R. B. Baculovirus Production of Fully-Active Phosphoinositide 3-Kinase alpha as a p85 α -p110 α Fusion for X-ray Crystallographic Analysis with ATP Competitive Enzyme Inhibitors. *Protein Expression Purif.* **2010**, 73, 167–176.